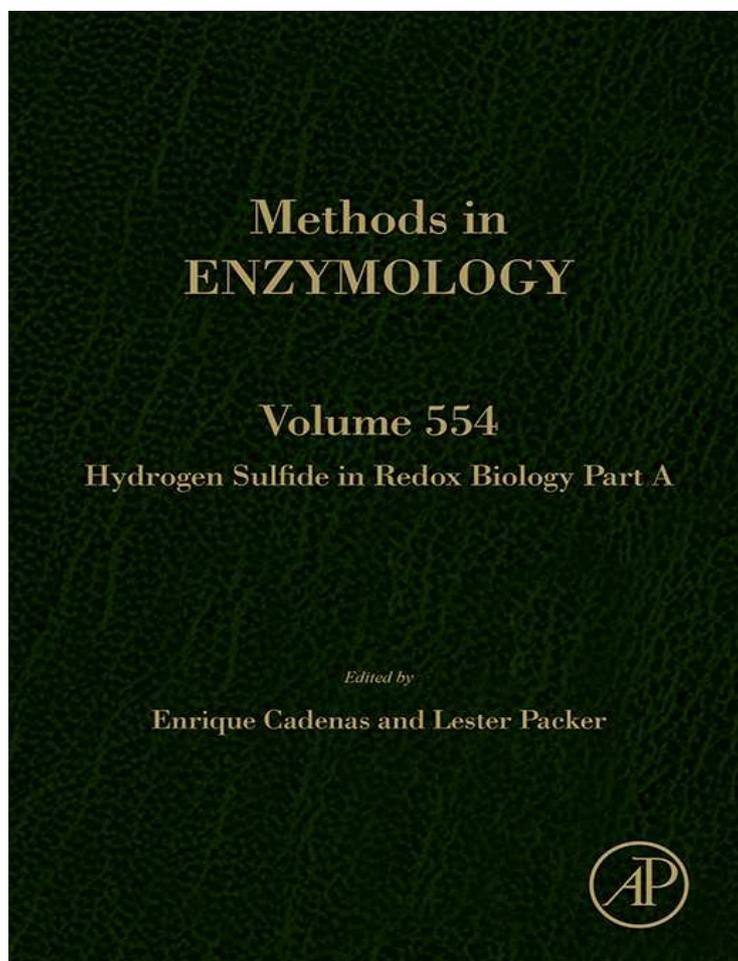


**Provided for non-commercial research and educational use only.  
Not for reproduction, distribution or commercial use.**

This chapter was originally published in the book *Methods in Enzymology, Vol. 554* published by Elsevier, and the attached copy is provided by Elsevier for the author's benefit and for the benefit of the author's institution, for non-commercial research and educational use including without limitation use in instruction at your institution, sending it to specific colleagues who know you, and providing a copy to your institution's administrator.



All other uses, reproduction and distribution, including without limitation commercial reprints, selling or licensing copies or access, or posting on open internet sites, your personal or institution's website or repository, are prohibited. For exceptions, permission may be sought for such use through Elsevier's permissions site at:

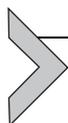
<http://www.elsevier.com/locate/permissionusematerial>

From Peter Rose, Brian W. Dymock and Philip K. Moore, GYY4137, a Novel Water-Soluble, H<sub>2</sub>S-Releasing Molecule. In: Enrique Cadenas and Lester Packer, editors, *Methods in Enzymology, Vol. 554*, Burlington: Academic Press, 2015, pp. 143-167.

ISBN: 978-0-12-801512-4

© Copyright 2015 Elsevier Inc.

Academic Press



# GY4137, a Novel Water-Soluble, H<sub>2</sub>S-Releasing Molecule

Peter Rose\*, Brian W. Dymock<sup>†</sup>, Philip K. Moore<sup>‡,1</sup>

\*University of Lincoln, Lincoln, Lincolnshire, United Kingdom

<sup>†</sup>Department of Pharmacy, National University of Singapore, Singapore

<sup>‡</sup>Neurobiology Program, Life Science Institute and Department of Pharmacology, National University of Singapore, Singapore

<sup>1</sup>Corresponding author: e-mail address: dprmpk@nus.edu.sg

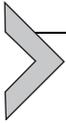
## Contents

1. Introduction	144
2. Why Slow Releasing H <sub>2</sub> S Donors?	146
3. The Development and Characterization of GY4137	147
4. Facile Synthesis and Chemical Characterization of GY4137	148
5. Biological Effects of GY4137: An Overview and Potential Role in Disease?	149
5.1 Cardiovascular system: Vascular smooth muscle and platelet function	152
5.2 Effect of GY4137 on nonvascular smooth muscle	153
5.3 Inflammation: Is GY4137 pro- or anti-inflammatory?	153
5.4 Effect of GY4137 in the reproductive system	155
5.5 GY4137: Apoptosis and cell cycle progression	156
5.6 GY4137 and aging	158
6. The Effect of GY4137 in Nonmammalian Systems	159
7. Conclusion	161
References	162

## Abstract

Hydrogen sulfide (H<sub>2</sub>S) is now recognized as the so called “third gasotransmitter” taking its place alongside nitric oxide and carbon monoxide. In recent years, H<sub>2</sub>S has been reported to exhibit a diverse range of pharmacological effects in biological systems. Much of this evidence is derived from a combination of conventional pharmacological and genetic approaches coupled with the use of chemical compounds such as sodium hydrosulfide, a rapid H<sub>2</sub>S releasing donor. Developments in the design of new drug entities which attempt to take into account physicochemical properties, targeting to specific cellular organelles, triggering of H<sub>2</sub>S release upon specific chemical reactions in the cell, and controlling the release of H<sub>2</sub>S over extended periods of time have been described. For most of these molecules, little or no work has been conducted to determine their biological activity or possible therapeutic effects. It is therefore not clear whether such molecules have therapeutic potential which highlights the need for further *in vivo* studies. One exception to the general rule is GY4137 (morpholin-4-ium

4-methoxyphenyl(morpholino) phosphinodithioate), a slow releasing H<sub>2</sub>S donor, which has been evaluated for activity in a range of pharmacological models both *in vitro* and *in vivo*. GYY4137 was first reported to release H<sub>2</sub>S and exhibit vasodilator activity over 5 years ago and, to date, GYY4137 is becoming increasingly employed as a pharmacological “tool” to explore the biological functions of H<sub>2</sub>S.



## 1. INTRODUCTION

In recent years much attention has been focussed on elucidating the functional roles of the gaseous mediator hydrogen sulfide (H<sub>2</sub>S) in biological systems (for reviews see, [Guo, Cheng, & Zhu, 2013](#); [Li, Rose, & Moore, 2011](#); [Olas, 2014](#); [Wang, 2012](#)). It is now clear that biologically active quantities of H<sub>2</sub>S are synthesized naturally by a number of cell types in a range of different tissues in mammals, nonmammalian species, and even plants. Moreover, evidence is growing that its principal role is to act as a signaling molecule bringing about a distinct set of physiological and biochemical effects.

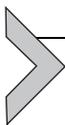
Considerable emphasis has been placed on determining the role of this gas in normal physiological processes and, at the same time, whether changes in the homeostatic regulation of the H<sub>2</sub>S system can trigger, contribute to, or exacerbate disease. A better understanding of the physiology and pathophysiology of H<sub>2</sub>S will be key to the development of novel therapeutics based on the general principles of either, (i) reducing endogenous H<sub>2</sub>S biosynthesis or, as will become apparent in this review, more likely, (ii) replenishing deficient production of this evanescent gas.

Much progress has been made in understanding the biology of H<sub>2</sub>S in the last decade. Thus, without going into specifics which will be dealt with elsewhere in this volume, H<sub>2</sub>S has been reported to play a part in a range of biologically fundamental processes which affect most cells/tissues including mitochondrial function, cytoprotection, inflammation, vascular perfusion, tissue preservation and repair, apoptosis, cell cycle regulation, and aging. Interestingly, many of these effects of H<sub>2</sub>S in biological systems are not restricted to mammalian species. For example, biosynthesis of this gas has been detected in the tissues from organisms found across several taxonomic kingdoms ([Calderwood & Kopriva, 2014](#); [Clarke, 1953](#); [Julian, Statile, Wohlgemuth, & Arp, 2002](#); [Olson et al., 2008](#); [Papenbrock, Riemenschneider, Kamp, Schulz-Vogt, & Schmidt, 2007](#)). In the plant

kingdom, exogenous application of H<sub>2</sub>S has been reported to increase crop yields, germination rates, influence root development, and alleviate heavy metal ion induced stress responses (Calderwood & Kopriva, 2014; Lisjak, Teklic, Wilson, Whiteman, & Hancock, 2013). Certainly, a considerable body of work now exists describing the functional roles of H<sub>2</sub>S in bacterial, animal, and plant systems.

Much attention has, not surprisingly, been focussed on the mechanism(s) by which cells generate H<sub>2</sub>S. By and large, the synthesis and production of this molecule is largely mediated *via* enzymatic routes. Recent evidence supports the liberation of H<sub>2</sub>S from endogenous persulfides and polysulfide species (Greiner et al., 2013; Ida et al., 2014; Miranda & Wink, 2014). Intracellular levels of glutathione persulfide were quantified at over 100 μM in mouse brain leading to the hypothesis that exogenous H<sub>2</sub>S is taken up primarily into the “persulfide pool.” The biological equilibrium is hence heavily weighted in favor of persulfides which could be major sulfur transfer signaling molecules. The major biosynthetic origins for H<sub>2</sub>S in mammals are *via* the catabolic activity of cystathione β synthetase (CBS, EC 4.2.1.22), cystathionine γ lyase (CSE, EC 4.4.1.1) on L-cysteine and 3-mercaptopyruvate sulfurtransferase (3-MST, EC 2.8.1.2) on 3-mercaptopyruvate. Worth noting is that additional systems are also likely involved in H<sub>2</sub>S homeostasis yet their relative contribution in maintaining tissue H<sub>2</sub>S levels remains largely unknown and, undoubtedly, other pathways responsible for H<sub>2</sub>S biosynthesis and degradation will likely be discovered.

Perturbations in endogenous H<sub>2</sub>S biosynthesis *via* one or other of these enzymatic pathways, may cause or play a part in disease states. For the most part, these disease states have been linked to loss of H<sub>2</sub>S production due to reduced activity or expression of CSE, CBS, 3-MST, or a combination thereof. Although somewhat simplistic, a variety of conditions including hypertension, inflammation, atherosclerosis, sexual dysfunction and reperfusion injury, and perhaps also cancer, are associated with reduced H<sub>2</sub>S generation or availability. Clearly, multiple pathologies and mechanisms underpin these diseases but, intriguingly, a lack of H<sub>2</sub>S production seems to be at least one common thread. Much effort has therefore been expended to understand the processes whereby both the activity and the expression of CSE, CBS, and 3-MST are controlled. This work is ongoing and will help to unravel the complex biological significance of H<sub>2</sub>S.

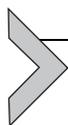


## 2. WHY SLOW RELEASING H<sub>2</sub>S DONORS?

As noted above it is becoming increasingly clear that a number of disease states have been linked to what might be described as a state of “relative H<sub>2</sub>S deficiency” in different cells, tissues and/or organs. As such, the expeditious use of H<sub>2</sub>S donor molecules may, logically, provide a possible common therapeutic approach to a wide range of disease states by replenishing H<sub>2</sub>S. A number of H<sub>2</sub>S donors have been reported in recent years. In addition to H<sub>2</sub>S gas itself, by far the most commonly used are the simple sulfide salts such as NaHS and Na<sub>2</sub>S. The use of these salts has been widespread in the literature over the last decade and has undoubtedly added greatly to our understanding of the complex biological effects of H<sub>2</sub>S. It could even be argued that the widespread availability of, for example, NaHS, which is both inexpensive to purchase and readily soluble in water, has been a crucial factor in creating the interest and giving impetus to researchers to study H<sub>2</sub>S, especially so in the early days. However, a major concern with NaHS is that it does not allow for the controlled release of H<sub>2</sub>S at rates that would be expected to mimic physiological conditions. Indeed, once solubilized, NaHS releases large amounts of H<sub>2</sub>S spontaneously, i.e., a bolus effect. It is very unlikely that endogenous H<sub>2</sub>S, made enzymatically in cells and tissues as detailed above, could ever be generated at the very high concentrations and over the very short time period, which is the case when cells or tissue are exposed to NaHS. One caveat in this argument is that the concentration of endogenous H<sub>2</sub>S generated enzymatically within individual tissues or indeed within individual subcellular components inside the target cell is not known and will require the development of much more sensitive and selective methods to detect this gas than are currently available.

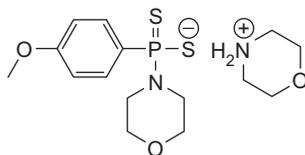
With this in mind, a consequence of the explosive H<sub>2</sub>S release profile of NaHS is that the biological effects elicited by this, or like, compounds when introduced into a biological system (i.e., a conscious or anesthetized animal, cells maintained in culture or tissues in organ baths) reflects the pharmacological (perhaps toxicological) activity of H<sub>2</sub>S and not its physiological role within that tissue/cell. As such, the use of compounds like NaHS, whilst valuable, may in some cases, obscure rather than illuminate, the biology of this gas. Clearly, using NaHS to probe for physiological roles of H<sub>2</sub>S is problematic. One approach to overcome this shortcoming has been the development of organic molecules which slowly release H<sub>2</sub>S at rates which are more akin to the rate expected from cells *in vivo* (Song et al., 2014).

A number of H<sub>2</sub>S donors of different types are now available. Many of these have been designed with the specific intent of delivering controlled levels of H<sub>2</sub>S gas to tissues. Of particular interest are a number of structurally divergent molecule classes encompassing H<sub>2</sub>S releasing nonsteroidal anti-inflammatory drugs (Isenberg et al., 2007; Sparatore et al., 2009), L-cysteine activated arylthioamide H<sub>2</sub>S donor molecules (Martelli et al., 2013), caged gem-dithiol derivatives that release H<sub>2</sub>S upon light stimulation (Devarie-Baez et al., 2013) and molecules that release H<sub>2</sub>S in aqueous systems. Many of these compounds, that are now available commercially, have been reviewed elsewhere (Kashfi & Olson, 2013; Song et al., 2014). More recently, additional H<sub>2</sub>S donors have been introduced including poly(ethylene glycol)-ADT (PEG-ADT) (Hasegawa & van der Vlies, 2014), and S-arylythiooximes (Foster, Powell, Radzinski, & Matson, 2014). In some cases, molecules which target delivery of H<sub>2</sub>S to specific organelles within cells, notably mitochondria, have been reported (Le Trionnaire, Perry, Whatmore, Wood, & Whiteman, 2013; Le Trionnaire et al., 2014). The advantage of all of these new drugs (c.f. NaHS) is their potential at least to provide a means to manipulate endogenous H<sub>2</sub>S levels within biological systems over a longer time course and within the range of concentrations likely to be achieved naturally. This approach will provide researchers with a wider range of “tools” to probe the biological significance of H<sub>2</sub>S and, perhaps in due course, will provide clinicians with alternative therapeutics.



### 3. THE DEVELOPMENT AND CHARACTERIZATION OF GYY4137

GYY4137 (morpholin-4-ium 4-methoxyphenyl (morpholino) phosphinodithioate) was described as a slow releasing H<sub>2</sub>S donor with vasorelaxant activity both *in vitro* and *in vivo* in 2008 (Li et al., 2008). Interestingly, GYY4137 was initially developed as a chemical entity for use as an accelerator in the vulcanization of natural rubber back in the late 1950s but it was not until several decades later that its use as a potential drug candidate was envisaged. GYY4137 was the first organic small molecule, to be reported and characterized as a slow releasing H<sub>2</sub>S donor (Fig. 1). GYY4137 is highly water soluble (>1 mg/mL at pH7.4), releases H<sub>2</sub>S at a slow rate (unlike sulfide salts) and is relatively easy to synthesize chemically or can be obtained commercially. To date, the biological effects of



**Figure 1** The chemical structure of GYY4137.

GYY4137 have been examined both *in vitro* and *in vivo* and a number of these studies will be described later in this article.

Initial work on the solution chemistry of GYY4137 indicated that it released low quantities of H<sub>2</sub>S over a sustained period (hours to days) in aqueous solution (pH 7.4, 37 °C), (Li et al., 2008). Moreover, the release of H<sub>2</sub>S from GYY4137 was pH and temperature dependent, with less H<sub>2</sub>S released at 4 °C and greater release at pH 3.0. Confirmation of the H<sub>2</sub>S releasing properties of GYY4137 within biological settings has also been demonstrated. Release of H<sub>2</sub>S from GYY4137 has now being independently verified using a range of assay procedures including use of the fluorescent probe 2,6-dansyl azide and 5,5'-dithiobis(2-nitrobenzoic acid) (Qabazard et al., 2014) alongside more refined procedures such as the H<sub>2</sub>S selective polarographic electrode (Kolluru, Shen, Bir, & Kevil, 2013) and detection of H<sub>2</sub>S using fluorescent HPLC analysis of monobromobimane derivatives (Shen, Peter, Bir, Wang, & Kevil, 2012).

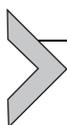
#### 4. FACILE SYNTHESIS AND CHEMICAL CHARACTERIZATION OF GYY4137

GYY4137 can be conveniently prepared by ring opening of Lawesson's Reagent (0.8 g, 2 mmol) with morpholine (0.87 g, 1 mmol) in dichloromethane (20 ml) solvent at room temperature. Upon stirring for about 2 h, the product slowly precipitates from the reaction mixture forming an ionic salt. This salt is filtered, washed with dichloromethane (10 ml, twice) to good purity. The solid is dried under vacuum at room temperature avoiding heat. GYY4137 is highly water soluble and care must be taken to avoid any aqueous work-up or washing with polar solvents. Once the solid is isolated, it is characterized *via* spectroscopic methods. Nuclear Magnetic Resonance (NMR) in D<sub>2</sub>O solvent confirms the identity and structure of GYY4137, (<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O) δ 8.02–7.84 (m, 2H), 7.05–6.94 (m, 2H), 3.93–3.84 (m, 4H), 3.81 (s, 3H), 3.71–3.54 (m, 4H), 3.23–3.21

(m, 4H), 2.86 (dd,  $J=9.6, 5.5$  Hz, 4H). <sup>13</sup>C NMR (101 MHz, D<sub>2</sub>O)  $\delta$  161.03 (d,  $J$  (C–P)=3.0 Hz), 132.48 (d,  $J$  (C–P)=12.8 Hz), 131.43 (d,  $J$  (C–P)=109.7 Hz), 113.56 (d,  $J$  (C–P)=14.6 Hz), 66.66 (d,  $J$  (C–P)=11.7 Hz), 63.64, 55.49, 44.72 (d,  $J$  (C–P)=1.8 Hz), 43.19. <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O)  $\delta$  89.00 (s). In the infrared spectrum a stretch at 650 cm<sup>-1</sup> can be seen for the P=S double bond.

GYY4137 should not be heated or exposed to moisture over time. It is recommended to store the solid material under nitrogen at -20 °C. For biological testing, a fresh solid sample should be weighed after allowing to warm to room temperature. For each assay, a solution in water or DMSO as required can be prepared and used freshly for the assay.

H<sub>2</sub>S release from GYY4137 can conveniently be quantified using a dansyl azide probe *in vitro* (Wang, Peng, Ni, Dai, & Wang, 2014). The recommended assay uses 2, 6-dansyl azide with a specific procedure in aqueous acetonitrile to ensure a good signal:noise ratio. Upon release of H<sub>2</sub>S from GYY4137 the sulfonyl azide group of the probe molecule rapidly reduces to a fluorescent primary sulfonamide product. Fluorescence accumulates over the time of the assay and can be measured using a plate reader at frequent time intervals up to days or even weeks. The limit of detection for this assay is around 1  $\mu$ M of released H<sub>2</sub>S. Concentrations of GYY4137 above 100  $\mu$ M, preferably incubated at 37 °C, are required to obtain a clear response.



## 5. BIOLOGICAL EFFECTS OF GYY4137: AN OVERVIEW AND POTENTIAL ROLE IN DISEASE?

A plethora of publications are now available describing one or other of the biological effects of NaHS in cells, tissues, and organs in health as well as in a range of experimentally induced disease states in animals. Any description of this body of work is beyond the scope of the current review but will be found elsewhere in this volume. In general, there is much less information available about the biological activity of any of the new classes of slow releasing H<sub>2</sub>S donors noted above and, indeed, for some of these compounds biological characterization is lacking altogether. At present, GYY4137 is without doubt the best studied of these H<sub>2</sub>S donors in biological terms and it is the findings from these studies that will be described here with an overview of the major biological effects of GYY4137 provided in Table 1.

**Table 1** Chronological summary of the major experimental evidence documenting the pharmacological effects of the novel H<sub>2</sub>S releasing molecule, GYY4137

Experimental model	Reported effect	Outcome	Reference
Rat aortic rings and perfused rat kidney <i>in vitro</i> and in the anesthetized rat <i>in vivo</i>	Vasorelaxant	Slow relaxation of rat aortic rings and dilated the perfused rat renal vasculature by opening vascular smooth muscle K <sub>ATP</sub> channels	Li et al. (2008)
Lipopolysaccharide (LPS; 4 mg/kg, i.v.) administered anesthetized rats	Anti-inflammatory	Inhibited LPS induced TNF- $\alpha$ production in rat blood. Decreased proinflammatory cytokines levels, TNF- $\alpha$ , IL-1 $\beta$ , IL-6, nitrite/nitrate, C-reactive protein, and L-selectin <i>in vivo</i>	Li, Salto-Tellez, Tan, Whiteman, and Moore (2009)
NO and H <sub>2</sub> S cross talk within the heart of mice	Contraction	Heart contractility induced by GYY4137, in the presence of the NO donor molecule sodium nitroprusside	Yong et al. (2011)
Anticancer activity in a murine xenograph model, subcutaneous transplantation of either HL-60 or MV4-11 cells	Inhibition of tumor growth	Administered daily—reduced tumor volume in HL-60 and MV4-11 injected animals	Lee et al. (2011)
A mouse model of acute joint inflammation	Anti-inflammatory	GYY4137 reduced synovial fluid myeloperoxidase (MPO) and <i>N</i> -acetyl- $\beta$ -D-glucosaminidase (NAG) activity and decreased TNF- $\alpha$ , IL-1 $\beta$ , IL-6, and IL-8 concentration with joints	Li et al. (2013) and Li, Yang, et al. (2013)
Platelet activation and microvascular thrombus formation	Antithrombotic activities	GYY4137 TRAP-induced adhesion molecule expression. Prolongation of venular thrombus formation and tail-vein bleeding time	Grambow et al. (2014)

---

High fat fed apolipoprotein E(-/-) mice	Antiatherosclerotic activity	Decreased aortic atherosclerotic plaque formation and partially restored aortic endothelium-dependent relaxation. Decreased ICAM-1, TNF- $\alpha$ , and IL-6 mRNA expression, and superoxide (O <sub>2</sub> ) generation in aorta. Increased aortic eNOS, phosphorylation expression of PI3K, enhanced Akt Ser(473) phosphorylation, and downregulated the expression of LOX-1.	<a href="#">Liu et al. (2013)</a>
High fat fed C57BL/6J mice	Prevention of insulin resistance	Decreased phosphorylation of perilipin 1 and hormone sensitive lipase	<a href="#">Geng et al. (2013)</a>
<i>In vitro</i> cell culture models across a range of immortalized and nonimmortalized cell lines	Anticancer properties	Exposure to NaHS or GYY4137 decreased cell survival more effectively in cancer cells. GYY4137, significantly increased glycolysis leading to overproduction of lactate, decreased anion exchanger, and sodium/proton exchanger activity. Combined effects leading to increased metabolic acid production, defective pH regulation, and cell death.	<a href="#">Lee et al. (2014)</a>

---

## 5.1. Cardiovascular system: Vascular smooth muscle and platelet function

One of the defining moments which spawned interest in the field of H<sub>2</sub>S biology was the finding that this gaseous molecule dilates blood vessels *in vitro* and *in vivo* (Hosoki, Matsuki, & Kimura, 1997). Since this discovery a number of the newly introduced H<sub>2</sub>S donor molecules, including GYY4137, have been found to exert similar vascular effects. The first report using GYY4137 identified a persistent and slowly developing relaxation of precontracted rat aortic rings which was antagonized by glibenclamide indicating that, like NaHS, this effect of GYY4137 is mediated *via* opening of vascular smooth muscle K<sub>ATP</sub> channels (Li et al., 2008). In a separate study, GYY4137 also relaxed precontracted mouse aortic rings (Bucci et al., 2012). In addition, GYY4137 dilated the perfused precontracted rat renal vasculature indicating that it acts not only on large capacitance vessels (e.g., aorta) but also on small resistance blood vessels which govern organ perfusion and blood pressure (Li et al., 2008). Finally, this report demonstrated that chronic treatment of rats with GYY4137 resulted in a lowering of blood pressure both in normotensive and, to a greater extent, in spontaneously hypertensive rats (Li et al., 2008). The effect of GYY4137 on cardiac motility has yet to be determined but one report did indicate that this H<sub>2</sub>S donor (like NaHS) could reduce the negative inotropic effect of NO (Yong et al., 2011). It should be noted whilst chronic treatment with GYY4137, using a dose regimen which reduced blood pressure for a period of several days, caused no apparent deleterious side effects in this study, a full screen for possible toxic side effects of GYY4137 has yet to be reported.

Apart from its effect on vascular smooth muscle, GYY4137 also has additional actions within the cardiovascular system. These include anti-thrombotic activity *via* inhibition of thrombin receptor agonist peptide (TRAP)-induced adhesion molecule expression and associated platelet activation (Grambow et al., 2014) as well as antiatherosclerotic activity in high fat fed, apolipoprotein E<sup>-/-</sup> mice (Liu et al., 2013). The latter study also showed pronounced effects of GYY4137 to inhibit expression not only of iNOS but also of P-selectin, lectin-like ox-LDL receptor-1, phosphorylated IκBα, NF-κB, ICAM-1, VCAM-1, and a range of chemokines including CXCL2, CXCR4, CXCL10, and CCL17 indicative of an anti-inflammatory effect which will be discussed in more detail later in this review. GYY4137 has also recently been reported to augment NO-mediated human platelet aggregation in response to collagen and

thrombin although the underlying mechanism is not yet clear (Ilkan et al., 2013). Similarly, and equally as important, is a potential role for H<sub>2</sub>S in regulating diabetic myocardial injury. In this context, high glucose concentrations promote cellular damage within cardiac tissues *via* the formation of advanced endpoint glycation products, increased reactive oxygen species (ROS) production and the generation of a proinflammatory environment. GYY4137 protects cardiac myoblast cells (H9c2) against high glucose-induced cytotoxicity by activation of the AMPK/mTOR signaling pathway. The activation of AMPK orchestrates the metabolic responses involved in the maintenance of appropriate energy levels needed for cell survival (Beauloye, Bertrand, Horman, & Hue, 2011; Wei, Hu, Zhuang, Liao, & Li, 2014). Such effects of GYY4137 may be related to a generalized action on cell metabolism which will be discussed below.

## 5.2. Effect of GYY4137 on nonvascular smooth muscle

A similar smooth muscle relaxant effect of GYY4137 has now been described in a range of tissues including human airway smooth muscle cells *via* opening of sarcolemma K<sub>ATP</sub> channels (Fitzgerald et al., 2014), mouse intrapulmonary airways by an effect on intracellular calcium release (Castro-Piedras & Perez-Zoghbi, 2013), pig bladder neck tissues again by opening K<sub>ATP</sub> channels (Fernandes et al., 2013), pregnant rat myometrial smooth muscle cells (Robinson & Wray, 2012) and in the bovine ciliary artery (Chitnis et al., 2013). Intriguingly, GYY4137 can occasionally, and in defined conditions, exert the opposite effect on blood vessels. Thus, in animals with experimentally induced endotoxic shock, GYY4137 reverses the hypotensive effect (i.e., increases blood pressure) due to *E coli* lipopolysaccharide (LPS) injection (Li et al., 2009). This likely reflects the anti-inflammatory effect of GYY4137 and, among other mechanisms, its ability to reduce the expression of inducible nitric oxide synthase (iNOS) which generates massive amounts of vasodilator nitric oxide (NO) in this condition.

## 5.3. Inflammation: Is GYY4137 pro- or anti-inflammatory?

GYY4137 exhibits anti-inflammatory properties in cultured cells such as macrophages *in vitro* and in a variety of animal models of inflammation *in vivo*. The anti-inflammatory properties ascribed to GYY4137 are typically concentration dependent with effects seen *in vitro* at concentrations ranging

from 0.05 to 5 mM and an effective dose administered intraperitoneally in mice of 50–100 mg/kg. In terms of inflammation, GYY4137 has been most extensively evaluated for its effect on cultured cells *in vitro*. In most cases, GYY4137 reduces the functionality of inflammatory cells by inhibiting NF- $\kappa$ B signaling in the target cells. Thus, GYY4137 pretreatment reduces the secretion of proinflammatory mediators like interleukin-1 $\beta$  (IL-1 $\beta$ ), interleukin-6 (IL-6), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), NO, and prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) (Whiteman et al., 2010) in mouse macrophages stimulated with LPS as well as in IL-8-stimulated airway smooth muscle cells (Perry et al., 2011). In human synoviocytes and articular chondrocytes, GYY4137 decreased LPS-evoked production of nitrite (NO<sub>2</sub><sup>-</sup>), PGE<sub>2</sub>, TNF- $\alpha$ , and IL-6 (Li, Fox, et al., 2013). Similarly, in human chondrocytes isolated from osteoarthritis tissues, GYY4137 reduces NO, PGE<sub>2</sub>, IL-6, and MMP-13 synthesis from cells stimulated with IL-1 $\beta$  (Fox et al., 2012). Stimulation of human chondrocytes isolated from osteoarthritis sufferers with IL-1 $\beta$  activates the NF- $\kappa$ B signaling pathway and the nuclear translocation and accumulation of this proinflammatory transcription factor. Induction of the NF- $\kappa$ B pathway is accompanied by increased production of the proinflammatory mediators PGE<sub>2</sub>, IL-6, and expression of proinflammatory proteins like MMP-13, cyclooxygenase-2 (COX-2), and iNOS, (Burguera, Vela-Anero, Magalhães, Meijide-Faílde, & Blanco, 2014). In this study, both GYY4137 and NaHS significantly reduced the levels of proinflammatory molecules and proteins in chondrocytes. An inhibitory effect of GYY4137 on NF- $\kappa$ B signaling has also been described in mouse macrophages infected with *Mycoplasma fermentans*. In this study, GYY4137 reduced the production of the proinflammatory cytokine MCP-1 and diminished nuclear accumulation of the NF- $\kappa$ B heterodimer p65/p52 (Benedetti et al., 2014).

GYY4137 also exhibits anti-inflammatory activity in animals *in vivo*. For example, injection of GYY4137 prior to LPS administration in the rat protects against the ensuing endotoxic shock both in terms of partially restoring the depressed blood pressure and with respect to inhibiting the generation of proinflammatory cytokines and NO (Li et al., 2009; Wang, Liu, et al., 2014). In addition, GYY4137 administered 18 h after intra-articular injection of Complete Freund's Adjuvant (CFA) in the mouse reduced paw swelling but when administered 1 h before CFA, GYY4137 increased swelling suggesting that the effect of this donor in this animal model critically depends on the stage of the inflammation. This data implies that H<sub>2</sub>S exerts different (perhaps even opposite) effects on the inflammatory response (Li, Fox, et al.,

2013; Li, Yang, Long, Yang, & Shen, 2013). In a recent report, GYY4137 coadministered with histamine into mouse skin inhibited the accompanying pruritis but did not affect the extravasations response (Rodrigues et al., 2013) again suggesting differences in the response to GYY4137 in different systems. Finally, administration of GYY4137 to newborn mouse pups subjected to hyperoxia (85% O<sub>2</sub> for 10 days) augmented lung alveoli development whilst blunting leuokocyte infiltration into the alveolar spaces indicative of an overall anti-inflammatory effect (Madurga et al., 2014). Cumulatively, these studies show that GYY4137 consistently reduces the expression and/or generation of proinflammatory mediators *in vitro* but has a more complex pattern of effects in animal models of inflammation *in vivo*. Inflammation is a multifactorial process which depends on the concerted and time-dependent interplay of numerous cell types each of which is able to generate a range of both pro- and anti-inflammatory mediators. Bearing in mind the somewhat promiscuous effect of H<sub>2</sub>S on different cells and transduction mechanisms it is perhaps not surprising that the *in vivo* effects of GYY4137 are complex. It is clear from *in vitro*, and from some studies *in vivo*, that the anti-inflammatory effect of GYY4137 is mediated by inhibiting NF-κB signaling. However, this by no means rules out other possible molecular targets for GYY4137 derived H<sub>2</sub>S which should continue to be sought to obtain a more complete picture of the mechanism of action of this H<sub>2</sub>S donor.

#### 5.4. Effect of GYY4137 in the reproductive system

The effect of GYY4137 has also been examined in the reproductive system. Roles for H<sub>2</sub>S in female reproductive biology have only recently come to light. For example, both CBS and CSE are expressed in human intrauterine tissues and placenta (Patel, Vatish, Heptinstall, Wang, & Carson, 2009; You et al., 2011) as well as in epithelial cells of the fallopian tubes (Ning et al., 2014). The first direct evidence suggesting a role for H<sub>2</sub>S in reproductive biology was in preeclampsia, a hypertensive condition that affects around 4% of all pregnancies. The administration of DL-propargylglycine (PAG, an inhibitor of the H<sub>2</sub>S synthesizing enzyme, CSE) to pregnant mice resulted in hypertension and the promotion of abnormal vascularization within the placenta. This was correlated with reduced placental growth factor production and increased soluble fms-like tyrosine kinase-1 (sFlt-1) and soluble endoglinin (sEng) release from human umbilical vein endothelial cells which are considered to be markers for the onset of preeclampsia in

humans. In mice, GYY4137 reversed this effect of PAG treatment, reducing circulating sFlt-1 and sEng levels and restoring fetal growth (Wang et al., 2013).

In the human fallopian tube, epithelium cell-derived H<sub>2</sub>S has been postulated to promote relaxation of the oviduct. Interestingly, egg retention within the fallopian tube is associated with ectopic pregnancy and deregulation of fallopian tube contractility may play a part in this process. Indeed, disruption of H<sub>2</sub>S signaling has recently been reported to promote embryo retention and cause developmental delay in the mouse whilst pharmacological treatment with either GYY4137 or NaHS reversed these effects and restored the status quo (Ning et al., 2014).

Although more work is clearly needed, these limited data do suggest possible roles for H<sub>2</sub>S in protecting against conditions such as preeclampsia and ectopic pregnancy and thereby raise the possibility that an H<sub>2</sub>S donor, like GYY4137, may be of value therapeutically in this regard.

## 5.5. GYY4137: Apoptosis and cell cycle progression

Whilst most reports in the literature have concentrated on the effect of GYY4137 in the cardiovascular system and in inflammation, it is worth noting that a potential use of this drug in the treatment of cancer has also recently begun to be addressed. To date, only a few reports have examined the involvement of H<sub>2</sub>S in cancer cell signaling and/or in the induction and promotion of apoptosis in cancer cells. From the available literature, a number of key findings have been made about the anticancer properties of H<sub>2</sub>S and donor molecules. Importantly, both pro- and anti-apoptotic effects of H<sub>2</sub>S have been reported perhaps related to the choice of H<sub>2</sub>S generating system used. For example, as noted earlier, conventional H<sub>2</sub>S donors such as NaHS and Na<sub>2</sub>S generate H<sub>2</sub>S rapidly in aqueous systems and the released H<sub>2</sub>S has a very short half life in cell culture media. It might be argued that GYY4137, which releases H<sub>2</sub>S slowly and continuously, may therefore be a better “tool” with which to evaluate the longer term effects of the gas in cell culture systems.

With this in mind, GYY4137 has been shown to influence cell cycle progression in a range of different cell types. This effect of GYY4137 appears to be cell type specific since it caused G2/M phase cell cycle arrest in immortalized MCF-7 cells (Lee et al., 2011), G1/S cell cycle transition in HepG2 cells (Lu, Gao, Huang, & Wang, 2014), and inhibits abnormal proliferation in lymphocytes derived from systemic lupus erythematosus

patients (Han et al., 2013). Aside from affecting cell cycle progression, GYY4137 also caused a concentration dependent loss of cell viability across a range of immortalized human cancer cell lines including HeLa, HCT-116, Hep G2, HL-60, MCF-7, MV4-11, and U2OS (Lee et al., 2011). Preliminary work has revealed that a number of GYY4137 chemical analogues exhibit similar activity (Liao et al., 2013). Mechanistic studies revealed that these effects were mediated *via* a caspase dependent pathway since PARP-1 and caspase 9 cleavage were evident in GYY4137-treated MCF-7 cells (Lee et al., 2011). The same study also reported that GYY4137 exerted a dose-dependent inhibitory effect on tumor growth in the mouse *in vivo*. Likewise, GYY4137 suppressed cell proliferation in human hepatocellular carcinoma (HCC) cell lines and reduced tumor growth *in vivo* most likely, in this case, by inhibition of the signal transducer and activator of transcription 3 (STAT3) pathway (Lu et al., 2014).

GYY4137 alters cell metabolism in a number of different ways some of which may have relevance in terms of its antitumor activity. For example, treatment of HeLa cells with GYY4137 increased calcium signaling by upregulating expression of the inositol 1,4,5-trisphosphate (IP<sub>3</sub>) receptors 1 and 2 (Lencesova et al., 2013). Moreover, depletion of calcium stores from the endoplasmic reticulum (ER) is accompanied by an increase in the expression of X-box, CHOP and ATF4, known biomarkers associated with ER stress (Lencesova et al., 2013). Similarly, upregulation of the sodium calcium exchanger (NCX) by GYY4137 elevates cAMP levels leading to increased expression of  $\beta_1$  and  $\beta_3$  adrenoceptors and the induction of apoptosis (Markova et al., 2013). These findings raise the possibility that GYY4137 may promote endoplasmic reticular stress *via* mobilization of calcium stores.

Historically, H<sub>2</sub>S gained notoriety due to its ability to inhibit the catalytic activity of cytochrome C oxidase, as H<sub>2</sub>S directly reduces the oxygen-reactive a<sub>3</sub>CuB binuclear center of this protein (Nicholls, Marshall, Cooper, & Wilson, 2013). It is perhaps no great surprise therefore that recent work has indicated subtle effects of GYY4137 on primary metabolic pathways within cancer cells. Aerobic glycolysis provides cancer cells with a growth or survival strategy as it provides valuable metabolic intermediates needed to support rapidly proliferating cells (Gatenby & Gillies, 2004). GYY4137 has recently been reported to inhibit glycolysis across a range of immortalized cancer cell lines with little or no effect on noncancer cells. Interestingly, overproduction of lactate and inhibition of the anion exchanger and sodium/proton exchanger lead to defective pH

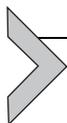
regulation, increased intracellular acidification, and cell death (Lee et al., 2014).

Clearly more work is needed to determine the role of extrinsic and intrinsic signaling pathways in the apoptotic effects of GYY4137 and to assess in greater detail its antitumor action *in vivo*. Moreover, the precise molecular mechanisms observed in these experimental studies also need to be examined.

## 5.6. GYY4137 and aging

Aging is intrinsically linked to many of the pathophysiological diseases described in this review. Any description of the biology of the aging process is beyond the scope of this review but readers will undoubtedly be aware of the profound and often deleterious effects that the aging process has on the cardiovascular system, metabolic functions, and immune system of mammalian species. Recently, it has been hypothesized that H<sub>2</sub>S plays a fundamental role in the aging process (Zhang et al., 2013). Evidence that H<sub>2</sub>S plays a role in this process stems from a number of observations notably, that H<sub>2</sub>S can inhibit oxidative stress (Benetti et al., 2013; Whiteman et al., 2004), increase longevity in some model organisms (Miller & Roth, 2007) and promote the induction of genes associated with longevity (Lee, Kennedy, Tolonen, & Ruvkun, 2003; Qabazard et al., 2014). H<sub>2</sub>S also reduces cell senescence and promotes cellular cytoprotective systems (Predmore, Alendy, Ahmed, Leeuwenburgh, & Julian, 2010). Indeed, these studies provided a potential approach to treat age-related deterioration in health. The big question is how does H<sub>2</sub>S influence the aging process and can GYY4137 be utilized as an antiaging drug in the sense of reducing the incidence or severity of age-related diseases? Recent evidence has indicated that GYY4137 can indeed reduce the deleterious effects of the aging process at least in the model nematode worm *Caenorhabditis elegans* (Qabazard et al., 2013, 2014). For example, GYY4137 treatment increased the expression of several age-related, stress response, and antioxidant genes. Moreover, GYY4137 increases lifespan in short-lived *mev-1* mutants with elevated oxidative stress and protected wild-type *C. elegans* against paraquat induced poisoning which is believed to occur *via* the generation of an excess of superoxide anions (Qabazard et al., 2013, 2014).

Taken together the available literature points to a fundamental role for H<sub>2</sub>S signaling in the aging process and provides rational avenues of intervention using H<sub>2</sub>S donor molecules like GYY4137.



## 6. THE EFFECT OF GYY4137 IN NONMAMMALIAN SYSTEMS

Whilst the majority of work highlighting a therapeutic role for GYY4137 has been derived from studies in mammalian models of disease less has been reported about the use of GYY4137 in other biological systems. This may be important since it provides further experimental evidence for biological roles for H<sub>2</sub>S and likely bolsters additional, albeit nonclinical, translational uses, for example to alleviate stress within agronomic crops.

To date, only a few reports have studied the effect of GYY4137 in non-mammalian species (for summary, see [Table 2](#)). For example, GYY4137 prevents Na<sup>+</sup> uptake in larval zebra fish, *Danio rerio* ([Kumai, Porteus, Kwong, & Perry, 2014](#)) and has antiaging properties when tested in the nematode worm, *C. elegans* ([Qabazard et al., 2013, 2014](#)). The effect of GYY4137 on plant metabolism has also been evaluated. Multiple reports over the years have shown that plants respond to H<sub>2</sub>S in a biphasic manner with both positive and negative effects on growth observed. From an environmental context, exposure of plants to high levels of exogenous sources of H<sub>2</sub>S gas can have phytotoxic effects with common symptoms associated with but, not restricted to, reduced oxygen production, reducing nutrient uptake, and the promotion of leaf damage ([Thompson & Kats, 1978](#)). In contrast, increased rates of photosynthesis, germination rates, and increased plant productivity upon exposure to H<sub>2</sub>S have also been documented ([Chen et al., 2011; Dooley, Nair, & Ward, 2013](#)). Interestingly, H<sub>2</sub>S production in plant tissues is likely derived from the sulfur assimilation pathways and L-cysteine desulfhydrase ([Ravilious & Jez, 2012; Romero et al., 2014; Takahashi, Kopriva, Giordano, Saito, & Hell, 2011](#)). It is known that in the presence of high concentrations of sulfate (SO<sub>4</sub><sup>2-</sup>), sulfur dioxide (SO<sub>2</sub>) and/or L-cysteine, plant tissues can synthesize, and emit H<sub>2</sub>S ([Hällgren & Fredriksson, 1982; Sekiya, Schmidt, Wilson, & Filner, 1982; Wilson, Bressan, & Filner, 1978](#)). Surprisingly, it was not until recently that a potential physiological role for H<sub>2</sub>S in plants was considered. To date, the reported effects of H<sub>2</sub>S on plants are wide ranging and this has led to many researchers postulating that H<sub>2</sub>S may function as a potential plant signaling molecule. Sadly, descriptions of potential roles for H<sub>2</sub>S releasing donor molecules in plants are rare. The use of H<sub>2</sub>S donor molecules should be considered in plant studies since H<sub>2</sub>S has the ability to promote plant growth and vigor, reduce the effect of biotic and abiotic stress and increase crop yields ([Dooley et al., 2013; Thompson, Kats, & Lennox, 1979](#)).

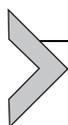
**Table 2** Studies utilizing GYY4137 to demonstrate novel physiological roles for H<sub>2</sub>S across a range of model species

Experimental model	Reported effect	Outcome	Reference
<i>Arabidopsis thaliana</i> ecotypes Landsberg erecta and Columbia ecotypes	Stomatal function	GYY4137 promotes stomatal opening in the light and prevented stomatal closure in the dark. Reduces the accumulation of NO within guard cells	Lisjak et al. (2010)
<i>Capsicum annuum</i>	Stomatal function	Reduced accumulation of NO induced by abscisic acid (ABA) treatment of leaf tissues. Promotes stomatal opening	Lisjak et al. (2011)
<i>Zea mays</i> L., Yedan no. 51	Prevention of heat damage	GYY4137 increased endogenous H <sub>2</sub> S levels, promoted survival, and reduced electrolyte leakage in seedlings under heat stress	Li, Fox, et al. (2013) and Li, Yang, et al. (2013)
<i>Capsicum annuum</i>	Prevention of salt stress	Accumulation of proline induced by salt treatment	Lisjak et al. (2013)
<i>Caenorhabditis elegans</i>	Aging	GYY4137 increased the expression of several age-related, stress response, and antioxidant genes. Increased lifespan in short-lived mev-1 mutants with elevated oxidative stress. Protected wild-type <i>C. elegans</i> against paraquat poisoning	Qabazard et al. (2014)
<i>Caenorhabditis elegans</i>	Survival and H <sub>2</sub> S homeostasis	GYY4137, extended median survival and increased tolerance toward oxidative and endoplasmic reticulum (ER) stress. Restored life span in cysl-2 mutant worms.	Qabazard et al. (2013)

Such studies highlighting a fundamental role of H<sub>2</sub>S in the regulation of aging, cellular stress pathways, and in plant physiological responses.

GYY4137 has already been used to explore the role of H<sub>2</sub>S in regulating stomatal function in plants (Lisjak et al., 2010, 2011). In these studies it was noted that H<sub>2</sub>S, derived either from NaHS or GYY4137, caused stomatal opening in the model plant *Arabidopsis thaliana* and in the commercially important crop species, *Capsium annuum*. Further experimentation revealed that both donors reduced abscisic acid induced accumulation of NO within leaf tissues suggestive of a potential cross talk between the two gases. Similarly, García-Mata and Lamattina (2010) also reported that GYY4137 promotes stomatal closure in *Vicia faba* (L.) var. *major* and in *Impatiens walleriana*.

The involvement of H<sub>2</sub>S, derived from GYY4137, in the prevention of plant stress responses has also recently been explored. In plants, exposure to high temperatures increases membrane fluidity and reduces membrane integrity, promotes protein denaturation, and reduces chloroplast and mitochondrial function leading to a state of oxidative and osmotic stress. In this regard, GYY4137 reduces the impact of heat stress and preserves the viability of *Zea Mays* (Li, Yang, et al., 2013). Moreover, preliminary data shows that GYY4137 induces expression of the cytoprotective amino acid proline in plant tissues following salt stress (Lisjak et al., 2013).



## 7. CONCLUSION

Much progress has been made in the characterization of the pharmacological effects of GYY4137 over the past several years. The biological activity of GYY4137 have now been comprehensively evaluated in a range of *in vitro* cell systems as well as in mammalian and nonmammalian species *in vivo* and also in plants. It is now clear that GYY4137 exhibits a very wide range of biological effects from vasodilator activity in isolated blood vessels through antiaging activity in *C. elegans* to crop preservation in plants. GYY4137 has the advantage that it is not only very water soluble but also easy to synthesize chemically as well as being commercially available. GYY4137 was an early addition to the family of H<sub>2</sub>S donors and can therefore be considered as the prototype for this class of compounds. Further progress will undoubtedly lead to new second and third generation compounds with the ability to release their H<sub>2</sub>S “payload” at well-defined rates, in defined organs and cells and in the appropriate subcellular location. Experimental “tools” of this type will prove invaluable in unraveling the complex biological roles of H<sub>2</sub>S in multiple systems and perhaps also act as the fore-runners of a new class of therapeutic agents.

## REFERENCES

- Beauloye, C., Bertrand, L., Horman, S., & Hue, L. (2011). AMPK activation, a preventive therapeutic target in the transition from cardiac injury to heart failure. *Cardiovascular Research*, *90*, 224–233.
- Benedetti, F., Davinelli, S., Krishnan, S., Gallo, R. C., Scapagnini, G., Zella, D., et al. (2014). Sulfur compounds block MCP-1 production by *Mycoplasma fermentans*-infected macrophages through NF- $\kappa$ B inhibition. *Journal of Translational Medicine*, *12*, 1–11.
- Benetti, L. R., Campos, D., Gurgueira, S. A., Vercesi, A. E., Guedes, C. E., Santos, K. L., et al. (2013). Hydrogen sulfide inhibits oxidative stress in lungs from allergic mice in vivo. *European Journal of Pharmacology*, *698*, 463–469.
- Bucci, M., Papapetropoulos, A., Vellecco, V., Zhou, Z., Zaid, A., Giannogonas, P., et al. (2012). cGMP-dependent protein kinase contributes to hydrogen sulfide-stimulated vasorelaxation. *PLoS One*, *7*, e53319.
- Burguera, E. F., Vela-Anero, A., Magalhães, J., Mejjide-Faílde, R., & Blanco, F. J. (2014). Effect of hydrogen sulfide sources on inflammation and catabolic markers on interleukin 1 $\beta$ -stimulated human articular chondrocytes. *Osteoarthritis and Cartilage*, *22*, 1026–1035.
- Calderwood, A., & Kopriva, S. (2014). Hydrogen sulfide in plants: From dissipation of excess sulfur to signalling molecule. *Nitric Oxide*. <http://dx.doi.org/10.1016/j.niox.2014.02.005>.
- Castro-Piedras, I., & Perez-Zoghbi, J. F. (2013). Hydrogen sulphide inhibits Ca<sup>2+</sup> release through InsP3 receptors and relaxes airway smooth muscle. *Journal of Physiology*, *591*, 5999–6015.
- Chen, J., Wu, F. H., Wang, W. H., Zheng, C. J., Lin, G. H., Dong, X. J., et al. (2011). Hydrogen sulphide enhances photosynthesis through promoting chloroplast biogenesis, photosynthetic enzyme expression, and thiol redox modification in *Spinacia oleracea* seedlings. *Journal of Experimental Botany*, *62*, 4481–4493.
- Chitnis, M. K., Njie-Mbye, Y. F., Opere, C. A., Wood, M. E., Whiteman, M., & Ohia, S. E. (2013). Pharmacological actions of the slow release hydrogen sulfide donor GYY4137 on phenylephrine-induced tone in isolated bovine ciliary artery. *Experimental Eye Research*, *116*, 350–354.
- Clarke, P. H. (1953). Hydrogen sulphide production by bacteria. *Microbiology*, *8*, 397–407.
- Devarie-Baez, N. O., Bagdon, P. E., Peng, B., Zhao, Y., Park, C. M., & Xian, M. (2013). Light-induced hydrogen sulfide release from “caged” gem-dithiols. *Organic Letters*, *15*, 2786–2789.
- Dooley, F. D., Nair, S. P., & Ward, P. D. (2013). Increased growth and germination success in plants following hydrogen sulfide administration. *PLoS One*, *8*, e62048.
- Fernandes, V. S., Ribeiro, A. S., Barahona, M. V., Orensanz, L. M., Martínez-Sáenz, A., Recio, P., et al. (2013). Hydrogen sulfide mediated inhibitory neurotransmission to the pig bladder neck: Role of K<sub>ATP</sub> channels, sensory nerves and calcium signalling. *Journal of Urology*, *190*, 746–756.
- Fitzgerald, R., DeSantiago, B., Lee, D. Y., Yang, G., Jae Yeon, K., Foster B, D., et al. (2014). H<sub>2</sub>S relaxes isolated human airway smooth muscle cells via the sarcolemmal K<sub>ATP</sub> channel. *Biochemical and Biophysical Research Communications*, *446*, 393–398.
- Foster, J. C., Powell, C. R., Radzinski, S. C., & Matson, J. B. (2014). S-arylothiooximes: A facile route to hydrogen sulfide releasing compounds with structure-dependent release kinetics. *Organic Letters*, *16*, 1558–1561.
- Fox, B., Schantz, J. T., Haigh, R., Wood, M. E., Moore, P. K., Viner, N., et al. (2012). Inducible hydrogen sulfide synthesis in chondrocytes and mesenchymal progenitor cells: Is H<sub>2</sub>S a novel cytoprotective mediator in the inflamed joint? *Journal of Cellular and Molecular Medicine*, *16*, 896–910.

- García-Mata, C., & Lamattina, L. (2010). Hydrogen sulphide, a novel gasotransmitter involved in guard cell signalling. *New Phytologist*, *188*, 977–984.
- Gatenby, R. A., & Gillies, R. J. (2004). Why do cancers have high aerobic glycolysis? *Nature Reviews Cancer*, *4*, 891–899.
- Geng, B., Cai, B., Liao, F., Zheng, Y., Zeng, Q., Fan, X., et al. (2013). Increase or decrease hydrogen sulfide exert opposite lipolysis, but reduce global insulin resistance in high fatty diet induced obese mice. *PLoS One*, *8*, e73892.
- Grambow, E., Mueller-Graf, F., Deliagina, E., Frank, M., Kuhula, A., & Vollmar, B. (2014). Effect of the hydrogen sulfide donor GYY4137 on platelet activation and microvascular thrombus formation in mice. *Platelets*, *25*, 166–174.
- Greiner, R., Palinkas, Z., Basell, K., Becher, D., Antelmann, H., Nagy, P., et al. (2013). Polysulfides link H<sub>2</sub>S to protein thiol oxidation. *Antioxidants & Redox Signaling*, *19*, 1749–1765.
- Guo, W., Cheng, Z. Y., & Zhu, Y. Z. (2013). Hydrogen sulfide and translational medicine. *Acta Pharmacologica Sinica*, *34*, 1284–1291.
- Hällgren, J. E., & Fredriksson, S. A. (1982). Emission of hydrogen sulfide from sulfur dioxide-fumigated pine trees. *Plant Physiology*, *70*, 456–459.
- Han, Y., Zeng, F., Tan, G., Yang, C., Tang, H., Luo, Y., et al. (2013). Hydrogen sulfide inhibits abnormal proliferation of lymphocytes via AKT/GSK3 $\beta$  signal pathway in systemic lupus erythematosus patients. *Cellular Physiology and Biochemistry*, *31*, 795–804.
- Hasegawa, U., & van der Vlies, A. J. (2014). Design and Synthesis of Polymeric Hydrogen Sulfide donors. *Bioconjugate Chemistry*, *25*, 1290–1300.
- Hosoki, R., Matsuki, N., & Kimura, H. (1997). The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant in synergy with nitric oxide. *Biochemical and Biophysical Research Communications*, *237*, 527–531.
- Ida, T., Sawa, T., Ihara, H., Tsuchiya, Y., Watanabe, Y., Kumagai, Y., et al. (2014). Reactive cysteine persulfides and S-polythiolation regulate oxidative stress and redox signalling. *Proceedings of the National Academy of Sciences of the United States of America*, *111*, 7606–7611.
- Ilkan, Z., Mustafa, F., Apostoli, G., Solomon, A., Whiteman, M., & Emerson, M. (2013). OP16 Hydrogen sulfide inhibits human platelet aggregation. *Nitric Oxide*, *31*(Suppl 2), S26. <http://dx.doi.org/10.1016/j.niox.2013.06.046>.
- Isenberg, J. S., Jia, Y., Field, L., Ridnour, L. A., Sparatore, A., Del Soldato, P., et al. (2007). Modulation of angiogenesis by dithiolethione-modified NSAIDs and valproic acid. *British Journal of Pharmacology*, *151*, 63–72.
- Julian, D., Statile, J. L., Wohlgemuth, S. E., & Arp, A. J. (2002). Enzymatic hydrogen sulfide production in marine invertebrate tissues. *Comparative Biochemistry and Physiology Part A, Molecular & Integrative Physiology*, *133*, 105–115.
- Kashfi, K., & Olson, K. R. (2013). Biology and therapeutic potential of hydrogen sulfide and hydrogen sulphide releasing chimeras. *Biochemical Pharmacology*, *85*, 689–703.
- Kolluru, G. K., Shen, X., Bir, S. C., & Kevil, C. G. (2013). Hydrogen sulfide chemical biology: Pathophysiological roles and detection. *Nitric Oxide*, *35*, 5–20.
- Kumai, Y., Porteus, C. S., Kwong, R. W., & Perry, S. F. (2014). Hydrogen sulfide inhibits Na<sup>+</sup> uptake in larval zebrafish, *Danio rerio*. *Pflügers Archiv/European Journal of Physiology*. <http://dx.doi.org/10.1007/s00424-014-1550-y>.
- Lee, S. S., Kennedy, S., Tolonen, A. C., & Ruvkun, G. (2003). DAF-16 target genes that control *C. elegans* life-span and metabolism. *Science*, *300*, 644–647.
- Lee, Z. W., Teo, X. Y., Tay, E. Y., Tan, C. H., Hagen, T., Moore, P. K., et al. (2014). Utilizing hydrogen sulfide as a novel anti-cancer agent by targeting cancer glycolysis and pH imbalance. *British Journal of Pharmacology*, *171*, 4322–43226.
- Lee, Z. W., Zhou, J., Chen, C. S., Zhao, Y., Tan, C. H., Li, L., et al. (2011). The slow-releasing hydrogen sulfide donor, GYY4137, exhibits novel anti-cancer effects in vitro and in vivo. *PLoS One*, *6*, e21077.

- Lencesova, L., Hudecova, S., Csaderova, L., Markova, J., Soltysova, A., Pastorek, M., et al. (2013). Sulphide signalling potentiates apoptosis through the up-regulation of IP3 receptor types 1 and 2. *Acta Physiologica*, 208, 350–361.
- Le Trionnaire, S., Perry, A., Szczesny, B., Szabo, C., Winyard, P. G., Whatmore, J. L., et al. (2014). The synthesis and functional evaluation of a mitochondria-targeted hydrogen sulfide donor, (10-oxo-10-(4-(3-thioxo-3H-1,2-dithiol-5-yl)phenoxy)decyl)triphenylphosphonium bromide (AP39). *Medicinal Chemistry Communications*, 5, 728–736.
- Le Trionnaire, S., Perry, A., Whatmore, J. L., Wood, M. E., & Whiteman, M. (2013). P50 Mitochondria-targeted hydrogen sulfide donors: A novel twist to an old “tail”? *Nitric Oxide*, 31(Suppl 2), S57. <http://dx.doi.org/10.1016/j.niox.2013.06.112>.
- Li, L., Fox, B., Keeble, J., Salto-Tellez, M., Winyard, P. G., Wood, M. E., et al. (2013). The complex effects of the slow releasing hydrogen sulfide donor GYY4137 in a model of acute joint inflammation and in human cartilage cells. *Journal of Cellular and Molecular Medicine*, 17, 365–376.
- Li, L., Rose, P., & Moore, P. K. (2011). Hydrogen sulfide and cell signaling. *Annual Review of Pharmacology and Toxicology*, 51, 169–187.
- Li, L., Salto-Tellez, M., Tan, C. H., Whiteman, M., & Moore, P. K. (2009). GYY4137, a novel hydrogen sulphide releasing molecule, protects against endotoxic shock in the rat. *Free Radical Biology and Medicine*, 47, 103–113.
- Li, L., Whiteman, M., Guan, Y. Y., Neo, K. L., Cheng, Y., Lee, S. W., et al. (2008). Characterization of a novel, water-soluble hydrogen sulfide-releasing molecule (GYY4137): New insights into the biology of hydrogen sulphide. *Circulation*, 117, 2351–2360.
- Li, Z. G., Yang, S. Z., Long, W. B., Yang, G. X., & Shen, Z. Z. (2013). Hydrogen sulphide may be a novel downstream signal molecule in nitric oxide-induced heat tolerance of maize (*Zea mays* L.) seedlings. *Plant, Cell and Environment*, 36, 1564–1572.
- Liao, L. X., Tsai, C. Y., Atan, M. S., Lee, Z. W., Deng, L. W., Dymock, B. W., et al. (2013). Identification of a novel slow releasing hydrogen sulfide donor for cancer therapy. *Nitric Oxide*, 31(Suppl 2), S47. <http://dx.doi.org/10.1016/j.niox.2013.06.090>.
- Lisjak, M., Srivastava, N., Teklic, T., Civale, L., Lewandowski, K., Wilson, I., et al. (2010). A novel hydrogen sulfide donor causes stomatal opening and reduces nitric oxide accumulation. *Plant Physiology and Biochemistry*, 48, 931–935.
- Lisjak, M., Teklic, T., Wilson, I. D., Whiteman, M., & Hancock, J. T. (2013). Hydrogen sulfide: Environmental factor or signalling molecule? *Plant, Cell and Environment*, 36, 1607–1616.
- Lisjak, M., Teklič, T., Wilson, I. D., Wood, M., Whiteman, M., & Hancock, J. T. (2011). Hydrogen sulfide effects on stomatal apertures. *Plant Signaling and Behaviour*, 6, 1444–1446.
- Liu, Z., Han, Y., Li, L., Lu, H., Meng, G., Li, X., et al. (2013). The hydrogen sulfide donor, GYY4137, exhibits anti-atherosclerotic activity in high fat fed apolipoprotein E(–/–) mice. *British Journal of Pharmacology*, 169, 1795–1809.
- Lu, S., Gao, Y., Huang, X., & Wang, X. (2014). GYY4137, a hydrogen sulfide (H<sub>2</sub>S) donor, shows potent anti-hepatocellular carcinoma activity through blocking the STAT3 pathway. *International Journal of Oncology*, 44, 1259–1267.
- Madurga, A., Mižíková, I., Ruiz-Camp, J., Vadász, I., Herold, S., Mayer, K., et al. (2014). Systemic hydrogen sulfide administration partially restores normal alveolarization in an experimental animal model of bronchopulmonary dysplasia. *The American Journal of Physiology - Lung Cellular and Molecular Physiology*, 306, L684–L697.

- Markova, J., Hudecova, S., Soltysova, A., Sirova, M., Csaderova, L., Lencesova, L., et al. (2013). Sodium/calcium exchanger is upregulated by sulfide signaling, forms complex with the  $\beta 1$  and  $\beta 3$  but not  $\beta 2$  adrenergic receptors, and induces apoptosis. *Pflügers Archiv European Journal of Physiology*, *466*, 1329–1342.
- Martelli, A., Testai, L., Citi, V., Marino, A., Pugliesi, I., Barresi, E., et al. (2013). Arylthioamides as H<sub>2</sub>S donors: l-cysteine-activated releasing properties and vascular effects in vitro and in vivo. *ACS Medicinal Chemistry Letters*, *4*, 904–908.
- Miller, D. L., & Roth, M. B. (2007). Hydrogen sulfide increases thermotolerance and lifespan in *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences of the United States of America*, *104*, 20618–20622.
- Miranda, K. M., & Wink, D. A. (2014). Persulfides and the cellular thiol landscape. *Proceedings of the National Academy of Sciences of the United States of America*, *111*, 7505–7506.
- Nicholls, P., Marshall, D. C., Cooper, C. E., & Wilson, M. T. (2013). Sulfide inhibition of and metabolism by cytochrome c oxidase. *Biochemical Society Transactions*, *41*, 1312–1316.
- Ning, N., Zhu, J., Du, Y., Gao, X., Liu, C., & Li, J. (2014). Dysregulation of hydrogen sulphide metabolism impairs oviductal transport of embryos. *Nature Communications*. <http://dx.doi.org/10.1038/ncomms5107>.
- Olas, B. (2014). Hydrogen sulfide in hemostasis: Friend or foe? *Chemico-Biological Interactions*, *217*, 49–56.
- Olson, K. R., Healy, M. J., Qin, Z., Skovgaard, N., Vulesevic, B., Duff, D. W., et al. (2008). Hydrogen sulfide as an oxygen sensor in trout gill chemoreceptors. *The American Journal of Physiology - Regulatory, Integrative and Comparative Physiology*, *295*, R669–R680.
- Papenbrock, J., Riemenschneider, A., Kamp, A., Schulz-Vogt, H. N., & Schmidt, A. (2007). Characterization of cysteine degrading and H<sub>2</sub>S releasing enzymes of higher plants - from the field to the test tube and back. *Plant Biology*, *9*, 582–588.
- Patel, P., Vatish, M., Heptinstall, J., Wang, R., & Carson, R. J. (2009). The endogenous production of hydrogen sulphide in intrauterine tissues. *Reproductive Biology and Endocrinology*, *7*, 1–9.
- Perry, M. M., Hui, C. K., Whiteman, M., Wood, M. E., Adcock, I., Kirkham, P., et al. (2011). Hydrogen sulfide inhibits proliferation and release of IL-8 from human airway smooth muscle cells. *The American Journal of Respiratory Cell and Molecular Biology*, *45*, 746–752.
- Predmore, B. L., Alendy, M. J., Ahmed, K. I., Leeuwenburgh, C., & Julian, D. (2010). The hydrogen sulfide signaling system: Changes during aging and the benefits of caloric restriction. *Age*, *32*, 467–481.
- Qabazard, B., Ahmed, S., Li, L., Arlt, V. M., Moore, P. K., Stürzenbaum, S. R., et al. (2013). *C. elegans* aging is modulated by hydrogen sulfide and the sulfhydrylase/cysteine synthase *cysl-2*. *PLoS One*, *8*, e80135.
- Qabazard, B., Li, L., Gruber, J., Peh, M. T., Ng, L. F., Kumar, S. D., et al. (2014). Hydrogen sulfide is an endogenous regulator of aging in *Caenorhabditis elegans*. *Antioxidant & Redox Signaling*, *20*, 2621–2630.
- Ravilious, G. E., & Jez, J. M. (2012). Structural biology of plant sulfur metabolism: From assimilation to biosynthesis. *Natural Products Reports*, *29*, 1138–1152.
- Robinson, H., & Wray, S. (2012). A New Slow Releasing, H<sub>2</sub>S Generating Compound, GYY4137 Relaxes spontaneous and oxytocin-stimulated contractions of human and rat pregnant myometrium. *PLoS One*, *7*, 1–12.
- Rodrigues, L., Santos, B. C., Florenzano, J., Dos Santos, K. T., Lopes, K. C., Lira, F. B., et al. (2013). P44 Does the slow-releasing hydrogen sulfide donor GYY4137 controls inflammation and pruritogen responses in mouse skin? *Nitric Oxide*, *31*(Suppl 2), S54–S55. <http://dx.doi.org/10.1016/j.niox.2013.06.106>.

- Romero, L. C., Aroca, M. Á., Laureano-Marín, A. M., Moreno, I., García, I., & Gotor, C. (2014). Cysteine and cysteine-related signaling pathways in *Arabidopsis thaliana*. *Molecular Plant*, 7, 264–276.
- Sekiya, J., Schmidt, A., Wilson, L. G., & Filner, P. (1982). Emission of hydrogen sulfide by leaf tissue in response to L-cysteine. *Plant Physiology*, 70, 430–436.
- Shen, X., Peter, E. A., Bir, S., Wang, R., & Kevil, C. G. (2012). Analytical measurement of discrete hydrogen sulfide pools in biological specimens. *Free Radical Biology & Medicine*, 52, 2276–2283.
- Song, Z. J., Ng, M. Y., Lee, Z.-W., Dai, W., Hagen, T., Moore, P. K., et al. (2014). Hydrogen sulfide donors in research and drug development. *Medicinal Chemistry Communications*, 5, 557–570.
- Sparatore, A., Perrino, E., Tazzari, V., Giustarini, D., Rossi, R., Rossoni, G., et al. (2009). Pharmacological profile of a novel H<sub>2</sub>S-releasing aspirin. *Free Radical Biology & Medicine*, 46, 586–592.
- Takahashi, H., Kopriva, S., Giordano, M., Saito, K., & Hell, R. (2011). Sulfur assimilation in photosynthetic organisms: Molecular functions and regulations of transporters and assimilatory enzymes. *Annual Review of Plant Biology*, 62, 157–184.
- Thompson, C. R., & Kats, G. (1978). Effects of continuous hydrogen sulfide fumigation on crop and forest plants. *Environmental Science & Technology*, 12, 550–553.
- Thompson, C. R., Kats, G., & Lennox, R. W. (1979). Effects of fumigating crops with hydrogen sulfide or sulphur dioxide. *Californian Agriculture*, 33, 9–10.
- Wang, R. (2012). Physiological implications of hydrogen sulfide: A whiff exploration that blossomed. *Physiological Reviews*, 92, 791–896.
- Wang, K., Ahmad, S., Cai, M., Rennie, J., Fujisawa, T., Crispi, F., et al. (2013). Dysregulation of hydrogen sulfide producing enzyme cystathionine  $\gamma$ -lyase contributes to maternal hypertension and placental abnormalities in preeclampsia. *Circulation*, 127, 2514–2522.
- Wang, C. N., Liu, Y. J., Duan, G. L., Zhao, W., Li, X. H., Zhu, X. Y., et al. (2014). CBS and CSE Are Critical for Maintenance of Mitochondrial Function and Glucocorticoid Production in Adrenal Cortex. *Antioxidants & Redox Signaling*. <http://dx.doi.org/10.1089/ars.2013.5682>.
- Wang, K., Peng, H., Ni, N., Dai, C., & Wang, B. (2014). 2,6-dansyl azide as a fluorescent probe for hydrogen sulphide. *Journal of Fluorescence*, 24, 1–5.
- Wei, W. B., Hu, X., Zhuang, X. D., Liao, L. Z., & Li, W. D. (2014). GYY4137, a novel hydrogen sulfide-releasing molecule, likely protects against high glucose-induced cytotoxicity by activation of the AMPK/mTOR signal pathway in H9c2 cells. *Molecular Cell Biochemistry*, 389, 249–256.
- Whiteman, M., Armstrong, J. S., Chu, S. H., Jia-Ling, S., Wong, B. S., Cheung, N. S., et al. (2004). The novel neuromodulator hydrogen sulfide: An endogenous peroxynitrite 'scavenger'? *Journal of Neurochemistry*, 90, 765–768.
- Whiteman, M., Li, L., Rose, P., Tan, C. H., Parkinson, D. B., & Moore, P. K. (2010). The effect of hydrogen sulfide donors on lipopolysaccharide-induced formation of inflammatory mediators in macrophages. *Antioxidants & Redox Signaling*, 12, 1147–1154.
- Wilson, L. G., Bressan, R. A., & Filner, P. (1978). Light-dependent Emission of Hydrogen Sulfide from Plants. *Plant Physiology*, 61, 184–189.
- Yong, Q. C., Cheong, J. L., Hua, F., Deng, L. W., Khoo, Y. M., Lee, H. S., et al. (2011). Regulation of heart function by endogenous gaseous mediators—crossstalk between nitric oxide and hydrogen sulfide. *Antioxidant & Redox Signaling*, 14, 2081–2091.

- You, X. J., Xu, C., Lu, J. Q., Zhu, X. Y., Gao, L., Cui, X. R., et al. (2011). Expression of cystathionine  $\beta$ -synthase and cystathionine  $\gamma$ -lyase in human pregnant myometrium and their roles in the control of uterine contractility. *PLoS One*, *6*, e23788.
- Zhang, Y., Tang, Z. H., Ren, Z., Qu, S. L., Liu, M. H., Liu, L. S., et al. (2013). Hydrogen sulfide, the next potent preventive and therapeutic agent in aging and age-associated diseases. *Molecular and Cellular Biology*, *33*, 1104–1113.