

1 **DNA methylation links prenatal smoking exposure to later life health outcomes in offspring**

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## 8 **Abstract**

9 **Background:** Maternal smoking during pregnancy is associated with adverse offspring health outcomes  
10 across their life course. We hypothesize that DNA methylation is a potential mediator of this relationship.

11 **Methods:** We examined the association of prenatal maternal smoking with offspring blood DNA  
12 methylation in 2,821 individuals (age 16 to 48 years) from five prospective birth cohort studies and perform  
13 Mendelian randomization and mediation analyses to assess, whether methylation markers have causal  
14 effects on disease outcomes in the offspring.

15 **Results:** We identify 69 differentially methylated CpGs in 36 genomic regions ( $P < 1 \times 10^{-7}$ ) associated with  
16 exposure to maternal smoking in adolescents and adults. Mendelian randomization analyses provided  
17 evidence for a causal role of four maternal smoking related CpG sites on an increased risk of inflammatory  
18 bowel diseases or schizophrenia. Further mediation analyses showed some evidence of cg25189904 in  
19 *GNG12* gene mediating the effect of exposure to maternal smoking on schizophrenia-related outcomes.

20 **Conclusions:** DNA methylation may represent a biological mechanism through which maternal smoking is  
21 associated with increased risk of psychiatric morbidity in the exposed offspring.

22 **Keywords:** maternal smoking, pregnancy, DNA methylation, persistence, mediation, disease, causality,  
23 lifecourse

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## 1 **Background**

2 Maternal smoking during pregnancy is associated with increased risk for pre-term birth, fetal growth  
3 restriction and low birth weight<sup>1-3</sup>, as well as neurodevelopmental impairments and respiratory and  
4 cardiovascular diseases later in life<sup>4-8</sup>. Despite these well-known risks, many women who commence  
5 pregnancy as smokers continue to smoke throughout gestation. According to a recent meta-analysis, the  
6 global prevalence of maternal smoking during pregnancy varies widely from a few percentages up to nearly  
7 40% in Ireland<sup>9</sup>. Thus, cigarette smoking continues to be one of the most important modifiable risk factors  
8 for the health of mothers and their children.

9  
10 Cigarette smoke is a potent environmental modifier of DNA methylation<sup>10</sup>. In support of this, an  
11 epigenome-wide meta-analysis of 13 birth cohort studies identified over 6,000 differentially methylated  
12 CpGs in cord blood of newborns exposed to prenatal smoking<sup>11</sup>. Several smaller studies have suggested  
13 that some of these methylation changes may persist across childhood and adolescence into adulthood<sup>12-15</sup>.  
14 However, questions remain concerning whether such DNA methylation changes endure across the life  
15 course and whether they play a mediating role in linking prenatal smoke exposure to later life health  
16 outcomes.

17  
18 Here, we combine data from five prospective birth cohort studies to investigate associations between  
19 prenatal smoking exposure and offspring blood DNA methylation in 2,821 adolescents and adults. We first  
20 examine the associations of prenatal smoking exposure with DNA methylation in each cohort and then  
21 meta-analyze the results across all studies. We focus on the >6,000 CpG sites previously identified in cord  
22 blood of newborns exposed to prenatal smoking<sup>11</sup>. We further (i) assess the impact of current smoking by  
23 the participant on DNA methylation; (ii) explore the dose-dependent effects of prenatal smoking exposure  
24 on methylation at key CpG sites; (iii) examine the potential intrauterine effect of smoking exposure on  
25 offspring DNA methylation by using paternal smoking as a negative control; (iv) assess the persistence of  
26 DNA methylation changes by investigating longitudinal associations from 30 to 48 years of age; and (v)

1 conduct Mendelian randomization (MR) and mediation analyses to examine the potential causal effects of  
2 DNA methylation changes on disease outcomes in the offspring (Figure 1). Our results show that prenatal  
3 smoking has persistent effects on the offspring epigenome and provide evidence for a causal role of DNA  
4 methylation in adverse health effects that may arise from exposure to tobacco smoke in utero.

5

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Insert Figure 1 here

7

## 8 **Results**

### 9 **Cohort specific characteristics of the study participants**

10 We analyzed the association of prenatal smoking exposure with blood DNA methylation in altogether 1,366  
11 adolescents (age 16y to 18y) and 1,455 adults (age 30y to 31y). Of these, 1,145 were from two independent  
12 Northern Finland Birth cohorts (NFBC 1966 and NFBC 1986), 257 were from the Isle of Wight Birth Cohort  
13 (IOWBC) and 1,419 from two Avon Longitudinal Study of Parents and Children cohorts (ALSPAC mothers  
14 and ALSPAC children). **Additional files 1-3** show the characteristics of each study cohort. Overall, 18.4% of  
15 the NFBC 1966 and 13.2% of the NFBC 1986 were prenatally exposed to maternal smoking. The  
16 corresponding figures were 11.8% for ALSPAC children, 28.7% for ALSPAC mothers and 16.3% for IOWBC.

17

### 18 **DNA methylation meta-analysis**

19 We found evidence for 69 differentially methylated CpGs in 36 genomic regions (**Table 1**). All of these CpG  
20 sites showed directionally concordant effects with previously reported associations in newborns<sup>11</sup> e.g.,  
21 hypermethylation of cg04180046 in *MYOG1* and cg05549655 in *CYP1A1* and hypomethylation of  
22 cg05575921 in *AHRR* and cg14179389 in *GFI1* in the exposed offspring compared with their unexposed  
23 counterparts.

24

25

Insert Table 1 here

26

1 **Sensitivity and downstream analyses**

2 To examine whether offspring's own smoking had influenced the results, we repeated the main analysis  
3 including only those individuals who had never smoked regularly in their life. The results were similar, in  
4 both direction and magnitude, across all 36 genomic regions as in the full meta-analysis (**Figure 2**),  
5 indicating that the association between maternal smoking and blood DNA methylation was not mediated  
6 through offspring's own smoking behavior.

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8 

9

10 We then examined the dose-response relationship between maternal smoking and blood DNA methylation  
11 in the offspring. Methylation differences between the exposed and unexposed offspring became larger  
12 with increased smoking intensity across most CpG sites, e.g. each additional three cigarettes smoked per  
13 day during pregnancy was associated with 0.23 standard deviation (SD) increase in methylation level in  
14 cg05549655 in *CYP1A1* gene (**Table 2**). **Figure 3** shows the visual representations of the dose-response  
15 effect of maternal smoking on offspring blood DNA methylation of top CpGs in four top loci.

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17 

18

19 To assess potential unmeasured confounding and to establish a causal intra-uterine effect between  
20 maternal smoking and the offspring DNA methylation we used paternal smoking as a negative control.  
21 Maternal smoking and paternal smoking showed similar directions of effect; however, the effect estimates  
22 for exposure to paternal smoking were considerably smaller (**Table 2**). Adjusting for paternal smoking had  
23 no significant effect on maternal smoking estimates (**Additional file 4**).

24

25 

26

1 We performed a longitudinal analysis to examine whether the maternal smoking associated alterations in  
2 DNA methylation persisted from early adulthood (age 30-31y) into midlife (age 46-48y) in the NFBC 1966  
3 and ALSPAC mothers cohorts. We found no evidence for change in direction or magnitude of associations in  
4 blood DNA methylation between the two time points (**Figure 4**), suggesting that DNA methylation levels  
5 remain relatively stable for several decades after prenatal exposure to maternal smoking.

6

7

Insert Figure 4 here

8

### 9 **Mendelian randomization analysis**

10 We estimated the causal effects of DNA methylation changes on disease outcomes using MR. We extracted  
11 the effect sizes of SNP-CpG associations for the 69 differentially methylated CpGs available in the Accessible  
12 Resource for Integrated Epigenomic Studies (ARIES) mQTL database<sup>16</sup> (<http://www.mqtl.db.org/>), and found  
13 strong instruments for 15 CpG sites. Of these 15 CpG sites, three (cg15578140 in microRNA 548f-3  
14 (*MIR548F3*), cg09935388 in Growth Factor Independent Protein 1 (*GFI1*), cg04598670 (unknown gene))  
15 showed potential causal associations with inflammatory bowel diseases and one (cg25189904 in Guanine  
16 Nucleotide Binding Protein Gamma 12 (*GNG12*)) with Schizophrenia ( $P_{FDR} < 0.05$ , **Table 3**).

17

18

Insert Table 3 here

19

### 20 **Mediation analysis**

21 We then sought to test whether methylation changes in these four CpGs mediated the association between  
22 maternal smoking and disease outcomes. However, since the prevalence of inflammatory bowel disease is  
23 relatively low in general population, we assessed the associations of maternal smoking and CpGs on  
24 irritable bowel syndrome (IBS), which is a constellation of functional gastrointestinal disorder symptoms.  
25 These data were obtained from self-administered questionnaires in NFBC1966 at 46 years<sup>17</sup>. Prevalence of  
26 schizophrenia is also low in the general population. Therefore, instead of diagnosed schizophrenia, we used

1 personality trait scales measuring schizotypal and affective symptoms as an outcome. Such personality  
2 scales were derived from questionnaires available in the NFBC 1966 data at 31 years and they can be used  
3 to identify subjects with latent personality with genetic vulnerability for schizophrenia<sup>18</sup>. We found  
4 evidence for cg25189904 mediating the association between exposure to maternal smoking and Bipolar II  
5 Scale ( $P = 0.024$ ) and Hypomanic Personality Scale ( $P = 0.018$ ) (**Figure 5 A and Figure 5 B**). The estimated  
6 mediated proportions were 30% and 28%, respectively (**Additional file 5**). We did not find evidence for a  
7 mediating effect of blood DNA methylation on IBS ( $P > 0.3$  for all CpGs, Additional file 5).

8  
9 Insert Figure 5 here  
10

## 11 **Discussion**

12 We combined data from five studies in adolescents and adults to examine the association between  
13 maternal smoking during pregnancy and blood DNA methylation in the offspring from 16 until 48 years of  
14 age. We identified 69 differentially methylated CpGs in 36 genomic regions. The top differentially  
15 methylated CpG sites showed a clear dose-response relationship with number of cigarettes smoked during  
16 pregnancy. The associations observed in adulthood were robust to adjustment for multiple potential  
17 confounding factors and persisted into middle age with no significant change in direction and magnitude of  
18 associations. Mendelian randomization and mediation analyses suggested that alterations in DNA  
19 methylation may link maternal smoking during pregnancy to increased risk of psychiatric morbidity and  
20 potentially with inflammatory bowel disease in the exposed offspring.

21  
22 The findings of our study confirm and extend the results of earlier reports by demonstrating that maternal  
23 smoking during pregnancy is associated with alterations in offspring blood DNA methylation not only in  
24 newborns<sup>11,19,20</sup>, children and adolescents<sup>12,13</sup>, but also in adults, several decades following the exposure.  
25 The similarity in differentially methylated CpG sites and the consistency in direction of methylation changes  
26 between our study and earlier EWAS imply that the smoke-exposure induced methylation changes may be

1 soma-wide and persist throughout life. However, the effects of smoking may also be targeted to specific  
2 regions of the epigenome, as indicated by the observations that both prenatal smoke exposure and active  
3 smoking affect the methylation patterns of same gene regions, e.g. *AHRR* and *CYP1A1*, which are involved  
4 in chemical detoxification<sup>10</sup>. Because of these similar effects, the methylation changes found in people  
5 exposed to prenatal smoking may also reflect current or past smoking by the people themselves or some  
6 other passive smoking exposure. Adjusting for offspring active smoking did not substantially change the  
7 results in the present study. However, parental smoking is known to associate with their offspring's  
8 smoking behavior also via genetic predisposition<sup>21,22</sup> and thus own smoking may serve as a mediator on the  
9 path between maternal smoking and DNA methylation. Therefore, simply adjusting for own smoking can  
10 lead to erroneous conclusions about the direct effects of maternal smoking<sup>23</sup>. We therefore performed a  
11 sensitivity analysis including only offspring who themselves had never smoked in their life and found that  
12 the associations were similar across all CpG sites as in the full meta-analysis.

13 We also used paternal smoking as a negative control by comparing the associations of maternal smoking  
14 during pregnancy and paternal smoking with offspring methylation and found that the effect estimates  
15 were substantially greater for maternal smoking, and adjusting for paternal smoking had virtually no effect  
16 on maternal smoking estimates. This indicates it is unlikely that the associations between maternal smoking  
17 and offspring methylation were attributable to post-natal passive smoking exposure or some unmeasured  
18 confounding. These results together with the finding of a clear dose-dependent relationship of methylation  
19 with increased smoking intensity during pregnancy suggest a direct biological effect of in utero exposure to  
20 cigarette smoke on DNA methylation.

21 The longitudinal analysis showed that differentially methylated CpGs observed around age 30 persisted into  
22 middle age (around age 48) without significant change in direction or magnitude of methylation levels. This  
23 corroborates the findings of recent smaller studies, which found several differently methylated CpGs in  
24 middle-aged women exposed to maternal smoking in utero<sup>14,15</sup> and suggests that some of the prenatal  
25 smoking exposure associated methylation changes are largely irreversible and unaffected by age and/or  
26 environmental exposures later in life. To assess whether such persistent changes in DNA methylation are

1 causally implicated with disease, we performed a Mendelian randomization analysis using summary data  
2 from large epigenome-wide association studies<sup>24</sup>. We found evidence for potential causal associations for  
3 three CpGs (cg15578140, cg09935388, cg04598670) with inflammatory bowel disease and one CpG  
4 (cg25189904) with schizophrenia. To strengthen the evidence for these potentially causal associations, we  
5 also performed a formal mediation analysis in the NFBC 1966 cohort and found evidence for differential  
6 methylation in cg25189904 mediating the association between maternal smoking and Bipolar II Scale and  
7 Hypomanic Personality Scale, explaining 30% and 28% of the total effect, respectively. These results  
8 corroborate the findings of previous observational studies, that maternal smoking during pregnancy is  
9 associated with increased risk of psychiatric morbidity in the exposed offspring<sup>25-28</sup>. However, we found no  
10 evidence for mediating effect of differential methylation cg15578140, cg09935388 and cg04598670 in the  
11 association of maternal smoking and irritable bowel syndrome. Such discrepant results could be due to  
12 relatively small sample size in the mediation analysis, or because the irritable bowel syndrome is not a good  
13 proxy for inflammatory bowel disease, or because the causal effect estimates for inflammatory bowel  
14 disease in the MR analysis were biased due to, for example, pleiotropic effects of genetic instruments on  
15 the outcome. Thus, additional studies are needed to assess whether prenatal smoking is associated with  
16 increased risk of inflammatory bowel disease in the exposed offspring and whether alterations in DNA  
17 methylation mediates this association.

18  
19 Our results may provide insights into potential mechanisms linking prenatal smoking exposure to  
20 psychiatric disorders. Experimental studies suggest that *GNG12* is an important regulator of inflammatory  
21 signaling in microglia cells, which are the resident macrophages of the central nervous system<sup>29</sup>. A role of  
22 inflammation in the etiology of schizophrenia and psychotic illness has been suggested<sup>30,31</sup>, and in line with  
23 this, a large meta-analysis of 2424 cases and over 1.2 million controls indicated that childhood central  
24 nervous system infections are associated with nearly two-fold risk of schizophrenia in adulthood<sup>32</sup>. Our  
25 DNA methylation data were from whole blood while the pathogenic processes for psychiatric disorders,  
26 including schizophrenia, occur primarily in brain tissue. We believe that methylation in blood mirrors the

1 corresponding sites in disease relevant tissues<sup>33</sup>. Such mirror sites can occur if the exposure occurs during  
2 early stages of prenatal development, thus affecting multiple tissues<sup>33</sup>. Therefore, blood DNA methylation  
3 may act as a marker for differential DNA methylation in the primary disease tissue that is mediating the  
4 effects of intrauterine smoke exposure. There is support justifying the use of blood samples to discover  
5 genes related to brain phenotypes and diseases<sup>34</sup>. However, further studies are needed to validate our  
6 findings and investigate the biological relevance of *GNG12* in the corresponding tissue.

7 Our study has both strengths and limitations. The large sample size of males and females and similar ages  
8 from different cohorts enabled us to obtain precise estimate of the long-term effects of maternal smoking  
9 on DNA methylation. Several downstream analyses and use of paternal smoking as a negative control  
10 allowed us to distinguish the associations from potential confounding, and the follow-up analysis from  
11 young adulthood to middle-age allowed us to examine the persistence of methylation changes. The  
12 limitations are that we did not have tissue-specific DNA methylation data as indicated above and that  
13 maternal smoking was determined from self-reported questionnaires. As self-reports may be biased by  
14 under-reporting or recall bias, our findings may be underestimate true effects. In the ALSPAC mothers  
15 cohort the adult offspring reported their mothers' smoking, although this could also be subject to recall  
16 bias. False reporting may also concern the adolescents in our study since they might have been reluctant to  
17 disclose their true smoking behavior, although in the IOWBC adolescent smoking was confirmed by urinary  
18 cotinine measurement. Another limitation is that the subjects in the ALSPAC children and ALSPAC mothers  
19 cohorts are related individuals. However, excluding either one of the related ALSPAC data sets did not  
20 notably affect the results (data not shown).

## 21 **Conclusions**

22 Maternal smoking during pregnancy has long-lasting effects on offspring epigenome. DNA methylation may  
23 represent a biological mechanism through which maternal smoking is associated with increased risk of  
24 psychiatric morbidity and potentially inflammatory bowel disease in the exposed offspring.

25

## 26 **Methods**

## 1 **Study cohorts**

### 2 Northern Finland Birth Cohort 1966

3 The Northern Finland Birth Cohort 1966, previously described in detail<sup>35,36</sup>, targeted all pregnant women,  
4 residing in the two northernmost provinces of Finland with expected dates of delivery between 1 January  
5 and 31 December 1966. Over 96 % of eligible women participated in the study, thus comprising 12,055  
6 mothers followed prospectively on average from 16<sup>th</sup> gestational week and 12,058 live born children. In  
7 1997, at offspring age of 31 years, all cohort participants with known addresses were sent a postal  
8 questionnaire on health and lifestyle and those living in Northern Finland or Helsinki area were invited to a  
9 clinical examination which included blood sampling. In total, both questionnaire and clinical data were  
10 collected for 6,007 participants. DNA was successfully extracted for 5,753 participants from fasted blood  
11 samples<sup>37</sup>. In 2012, all individuals with known address in Finland were sent postal questionnaires and an  
12 invitation for clinical examination. Both questionnaire and clinical data was collected for 5,539 participants.  
13 DNA methylation at 31 years was extracted for 807 randomly selected subjects of whom both  
14 questionnaire and clinical data with cardio-metabolic measures were available at both 31 and 46 years. Of  
15 these individuals, DNA methylation data at 46 years was extracted for 766 subjects.

### 16 Northern Finland Birth Cohort 1986

17 The Northern Finland Birth Cohort 1986 includes all mothers (prospective data collection from 10<sup>th</sup>  
18 gestational week) with children whose expected date of delivery fell between July, 1st 1985 and June, 30th  
19 1986, in the two northernmost provinces of Finland (99% of all births during that time)<sup>38</sup>. The cohort  
20 consists of 9,362 women and 9,432 live-born children. In 2001, all individuals with known address received  
21 a postal questionnaire on health and lifestyle and invitation to a clinical examination. DNA were extracted  
22 from fasting blood samples and DNA methylation were measured for 546 randomly selected subjects with  
23 full data available.

1 In both NFBC cohorts, complete data included singleton births and subjects with complete set clinical  
2 follow-up and DNA methylation data; excluding subjects with missing information and twins. A written  
3 informed consent for the use of the data including DNA was obtained from all study participants and their  
4 parents. Ethical approval for the study was received from Ethical Committee of Northern Ostrobothnia  
5 Hospital District and Oulu University, Faculty of Medicine.

#### 6 Isle of Wight Birth Cohort

7 Isle of Wight Birth cohort is a general population based birth cohort recruited on the Isle of Wight in 1989  
8 to assess the role of heredity and environment on development of allergic disorders and allergen  
9 sensitization. The details of this birth cohort have been described in previous reports<sup>39</sup>. In brief, both the  
10 Isle of Wight and the study population are 99% Caucasian. Ethics approvals were obtained from the Isle of  
11 Wight Local Research Ethics Committee (now named the National Research Ethics Service, NRES Committee  
12 South Central –Southampton B) at recruitment and for the 1, 2, 4, 10 and 18 years follow-up. Exact age at  
13 18-year follow-up was calculated from the date of blood sample collection for the 18-year follow-up and  
14 the date of birth. DNA methylation in peripheral blood samples was analyzed from randomly selected  
15 subjects (n = 257) at the 18-year follow-up.

#### 16 Avon Longitudinal Study of Parents and Children

17 Pregnant women resident in the former county of Avon, UK with expected dates of delivery 1st April 1991  
18 to 31st December 1992 were invited to take part in the study. The initial number of pregnancies enrolled is  
19 14,541 (for these at least one questionnaire has been returned or a “Children in Focus” clinic had been  
20 attended by 19/07/99). Of these initial pregnancies, there was a total of 14,676 fetuses, resulting in  
21 14,062 live births and 13,988 children who were alive at 1 year of age<sup>40,41</sup>.

22 The Accessible Resource for Integrated Epigenomic Studies (ARIES) is a sub study of ALSPAC, which includes  
23 1,018 mothers and their children for whom methylation data has been created<sup>42</sup>. The ARIES participants  
24 were selected based on the availability of DNA samples at two time points for the women (antenatal [mean

1 age 30 years] and at follow-up [mean age 48 years] when the offspring were adolescents) and three time  
2 points for their offspring (neonatal, childhood [mean age 7.5 years], and adolescence [mean age 17.1  
3 years]). A web portal allows openly accessible browsing of aggregate ARIES DNA methylation data (ARIES-  
4 Explorer) (<http://www.ariesepigenomics.org.uk/>). Please note that the study website contains details of all  
5 the data that is available through a fully searchable data dictionary and variable search tool:  
6 <http://www.bristol.ac.uk/alspac/researchers/our-data/>. Ethical approval for the study was obtained from  
7 the ALSPAC Ethics and Law Committee and the Local Research Ethics Committees.

### 8 **Definition of maternal smoking during pregnancy**

9 In NFBCs and ALSPAC studies, expectant mothers were asked whether they had smoked cigarettes before  
10 or at the beginning of the pregnancy, how many years they had smoked, the number of cigarettes smoked  
11 per day, and whether they had changed their smoking habits during the pregnancy. Offspring were  
12 considered to be prenatally exposed to cigarette smoking if mother reported smoking regularly (at least  
13 one cigarette per day) from pregnancy week 8 onwards. The ALSPAC mothers were also asked whether  
14 their mothers had smoked, and were asked whether they had smoked when they were pregnant with  
15 them. In the IOWBC, maternal smoking status in pregnancy was self-reported and defined as any smoking  
16 in pregnancy or no smoking during pregnancy.

### 17 **Measurement of DNA methylation**

18 Methylation of genomic DNA was quantified using the Illumina HumanMethylation450 array (ALSPAC,  
19 ARIES, IOWBC, and NFBC1966 at age 31, NFBC1986) or Illumina EPIC array (NFBC1966 at age 46) according  
20 to manufacturer's instructions. Bisulfite conversion of genomic DNA was performed using the EZ DNA  
21 methylation kit according to manufacturer's instructions (Zymo Research, Orange, CA).

22

### 23 **Quality control of methylation data**

1 In NFBCs and IOWBC quality control and quantile normalization for DNA methylation data was adapted  
2 from the CPACOR pipeline<sup>43</sup>. Illumina Background Correction was applied to the intensity values, a  
3 detection *P* value threshold was set at  $P < 10^{-16}$ , and samples with call rate < 98 % were excluded. Quantile  
4 normalization was done separately for six probe-type categories, and these normalized intensity values  
5 were used to calculate the methylation beta value at each CpG site, ranging between 0 (no methylation)  
6 and 1 (full methylation). Probes with call rate < 95 % were excluded from the analyses. A principal  
7 component analysis (PCA) was carried out for array control probes, and the first 30 principal components  
8 (PCs) were used as explanatory variables in the subsequent regression models<sup>43</sup>. White blood cell  
9 subpopulation estimates were obtained using the software provided by Houseman et al.<sup>44</sup> and these  
10 estimates were also added as covariates in the regression models. In ARIES, the DNA methylation wet-  
11 laboratory and pre-processing analyses were performed as previously described<sup>42</sup>. In brief, samples from  
12 all time points were distributed across slides using a semi-random approach to minimize the possibility of  
13 confounding by batch effects. Samples failing quality control (average probe *P* value  $\geq 0.01$ , those with sex  
14 or genotype mismatches) were excluded from further analysis and scheduled for repeat assay, and probes  
15 that contained <95% of signals detectable above background signal (detection *P* value <0.01) were  
16 excluded from analysis. Methylation data were pre-processed using R software, with background correction  
17 and subset quantile normalization performed using the pipeline described by Touleimat and Tost<sup>45</sup>.

## 18 **Statistical analyses**

### 19 **Meta-analysis of 6073 CpG sites in five studies**

20 Study design and analytical flow of the study are shown in **Figure 1** and the data availability for each  
21 analysis is presented in **Table 4**. All analyses were conducted using R software<sup>46</sup>. Linear regression was used  
22 to examine the association between sustained maternal smoking during pregnancy (from pregnancy week  
23 8 onwards) and offspring peripheral blood DNA methylation at 6073 CpG sites that were previously  
24 identified to be differentially methylated in newborns exposed to maternal smoking in utero in recent  
25 epigenome-wide association study (EWAS) (false discovery rate corrected *P* value < 0.05)<sup>11</sup>. The final model

1 was adjusted for study specific covariates as necessary (offspring's sex, BMI, smoking status and social class  
2 for IOWBC; additionally first four genetic PCs for NFBC cohorts; offspring age, maternal age and social class  
3 for ALSPAC cohorts). The model was run independently in each study, and the results were then meta-  
4 analyzed over all five studies (NFBC1986, NFBC1966 (age 16y & 31y), IOWBC (age 18y) and ALSPAC mothers  
5 (age 30y), ALSPAC children (age 17y) using an inverse-variance weighted fixed-effects model. Statistical  
6 significance level was set at  $P < 1 \times 10^{-7}$ , which corresponds approximately to a Bonferroni-corrected  
7 significance level of 0.05 for 450,000 independent tests. Such a conservative threshold was robust and thus  
8 the significant probes were considered worthy of further examination in a series of sensitivity and  
9 downstream analyses. The leading CpG site from each gene region (1 Mb window centered on the CpG site  
10 with the strongest association) was selected for these analyses.

11 We note that ALSPAC children were part of the earlier study from where the 6073 CpG sites were  
12 selected<sup>11</sup>. However, the earlier study examined DNA methylation in cord blood, whereas the current study  
13 uses blood DNA methylation data from the same cohort at 17 years. If the associations with exposure to  
14 maternal smoking in cord blood DNA methylation were due to confounding, we would not expect the signal  
15 to persist until adolescence. Furthermore, removal of the ALSPAC children from the meta-analysis made no  
16 material difference to the effect size estimates (data not shown).

17 Insert Table 4 here

## 18 **Sensitivity analyses**

### 19 ***Impact of offsprings' own smoking on their DNA methylation***

20 To assess the impact of participants' own smoking on methylation level by maternal smoking exposure, the  
21 same regression model was run excluding all participants who reported smoking regularly, defined in  
22 NFBC1966 as smoking at least one cigarette per day for one year or more during their life. In the ALSPAC  
23 mothers' cohort smoking behavior was queried at two time points. At age 30y, women were asked whether  
24 they had smoked regularly before pregnancy. At age 48y, women were asked whether they were current or

1 former smokers, and in case of the latter, whether they had smoked every day. From these data a  
2 dichotomous variable for any smoking for each of the time points was derived. In the IOWBC, participant's  
3 own smoking status was defined as having ever or never smoked asked via questionnaire administered at  
4 age 18y. The model was run independently in each study with the same covariates as above (excluding  
5 adjustment for offspring's smoking as all individuals were non-smokers) and meta-analyzed using an  
6 inverse-variance weighted fixed-effects model.

### 7 ***Impact of mother's smoking intensity on offspring DNA methylation***

8 Further analyses were performed to investigate whether intensity of maternal smoking during pregnancy  
9 had differential impact on the level of offspring blood DNA methylation. For this, the association between  
10 the number of cigarettes smoked per day during pregnancy and offspring blood DNA methylation was  
11 assessed in the NFBC studies. The association with the number of cigarettes smoked and offspring blood  
12 DNA methylation was assessed using linear regression with the same covariates as in the main analysis and  
13 meta-analyzed using an inverse-variance weighted fixed-effects model.

### 14 ***Negative control design to distinguish intrauterine effects from confounding***

15 Potential unmeasured confounding was examined in the NFBC studies by using paternal smoking status  
16 during pregnancy as a negative control. This method compares the associations of maternal and paternal  
17 smoking during pregnancy with offspring methylation outcomes. Use of paternal smoking as a negative  
18 control is based on the assumption that the biological effects of paternal smoking on intrauterine exposure  
19 are negligible compared to the effects of maternal smoking during pregnancy. If there is an intrauterine  
20 effect of cigarette smoke exposure, the associations are expected to be stronger for maternal smoking than  
21 paternal smoking behavior. If effects are of similar magnitude, the associations between maternal smoking  
22 during pregnancy and offspring methylation are likely attributable to unmeasured confounding, either by  
23 shared environmental or genetic factors<sup>47</sup>. The association with exposure to paternal smoking and offspring  
24 blood DNA methylation was assessed using linear regression with the same covariates as in the main  
25 analysis and meta-analyzed using an inverse-variance weighted fixed-effects model.

1 ***Persistence of DNA methylation into adulthood***

2 We also examined whether the methylation changes associated with maternal smoking persisted into  
3 middle-age. DNA methylation data were available at two time-points in NFBC 1966 (age 31y and 46y) and  
4 ALSPAC mother (age 30y and 48y). Generalized least squares were used to examine the longitudinal change  
5 in association between exposure to maternal smoking and blood DNA methylation. DNA methylation at  
6 each time point was regressed on the technical and white blood cell covariates, and the corresponding  
7 residuals were used as the outcome. Study-specific covariates (offspring sex, smoking, BMI and social class  
8 at each time point in NFBC1966; maternal age, social class and offspring age and smoking status at each  
9 time point in ALSPAC) were added in the model. Time point of measurement and its interaction with the  
10 exposure were added as additional terms to the regression model, and the model residuals were allowed to  
11 be correlated within each individual and be heteroskedastic between time points. The effect estimates at  
12 both time points can be derived from this model, and the test for equality of the estimates at both time  
13 points is equivalent to testing the interaction term being equal to zero<sup>48</sup>. The analyses were conducted  
14 separately in NFBC1966 and ALSPAC mothers and meta-analyzed using an inverse-variance weighted fixed-  
15 effects model.

16 ***Mendelian randomization analysis for the effect of DNA methylation on disease outcomes***

17 We next sought to assess the potential causal relationship between DNA methylation as the exposure and  
18 106 different diseases as outcomes available through the MR-Base platform (available at  
19 <http://www.mrbase.org/>) using two-sample Mendelian randomization (MR). The two-sample MR approach  
20 uses gene-exposure and gene-outcome associations from different data sources of comparable populations  
21 and allows the interrogation of summary estimates available from large genome-wide association study  
22 (GWAS) consortia<sup>24</sup>. If instrumental variable assumptions for the genes associated with the exposure are  
23 fulfilled<sup>49</sup>, then MR estimates can give evidence for a causal effect of exposure on the outcome.

24 We first looked up proxy single nucleotide polymorphisms (SNPs) for each of the 69 top maternal-smoking  
25 associated CpG sites in the publicly available ARIES database containing methylation quantitative trait loci

1 (mQTL) at four different life stages (birth, childhood, adolescence, middle age) in human blood<sup>42</sup>. We  
2 selected SNPs associated with each CpG at  $P < 10^{-7}$  at any of the other four time points. After clumping  
3 SNPs (using 1 Mb window and  $R^2 < 0.001$ ) and pruning the CpG sites to one per locus, we found strong  
4 instruments for 15 CpG sites (**Additional file 6**). These SNP-CpG associations were consistent across all time  
5 points (**Additional file 7**), except rs4306016-cg01825213 association, which was excluded from the final MR  
6 analysis. We selected the SNP-CpG and SNP-disease effects sizes at middle age and aligned these to the  
7 same allele. MR effect estimates were then calculated using Wald ratio or, in case of cg04598670, which  
8 had two SNP instruments available, inverse-variance weighted method. The resulting effect estimate  
9 represents the change in outcome per unit increase in the exposure.

#### 10 ***Mediation analysis***

11 The CpGs that showed evidence for causal relationship with disease outcomes in the MR analysis were  
12 tested for mediation in the association between maternal smoking during pregnancy and disease outcomes  
13 using the NFBC1966 data at 31y and 46y. We performed model-based causal mediation analysis using R  
14 package 'mediation'<sup>50</sup> by first estimating both the effect of maternal smoking on the CpG site and the effect  
15 of CpG site on the outcome, adjusted for exposure to maternal smoking (**Figure 6**). Both of these effects  
16 were additionally adjusted for sex, offspring's own smoking and technical covariates. We generated the  
17 estimates for the total effect, average direct effect and average causal mediation effect using quasi-  
18 Bayesian Monte Carlo method based on normal approximation with 2000 simulations, with robust standard  
19 errors. The proportion that the mediating CpG explains of the association between maternal smoking and  
20 disease outcome was calculated as described<sup>51</sup>.

21

22

#### 23 **Additional files**

1 Additional file 1. Characteristics of the participants based on exposure to maternal smoking during  
2 pregnancy in the NFBC cohorts (DOCX, 15 kb).

3 Additional file 2. Characteristics of the participants based on exposure to maternal smoking during  
4 pregnancy in the ALSPAC studies (DOCX, 14 kb).

5 Additional file 3. Characteristics of the participants based on exposure to maternal smoking during  
6 pregnancy in the IOWBC (DOCX, 13 kb).

7 Additional file 4. Paternal smoking adjusted association results of exposure to maternal smoking during  
8 pregnancy and offspring peripheral blood DNA Methylation for the top CpG sites (DOCX, 15 kb).

9 Additional file 5. Mediation analysis examining the mediated effect of maternal smoking during pregnancy  
10 on schizophrenia-related personality traits and inflammatory bowel syndrome in the NFBC 1966 cohort  
11 (DOCX, 13 kb).

12 Additional file 6. CpG site and their association with methylation in the ARIES cord blood data (DOCX, 15  
13 kb).

14 Additional file 7. Effect sizes and their 95% confidence intervals of each available SNP-CpG association  
15 across different time point in the ARIES data (DOCX, 109 kb).

16

## 17 **Declarations**

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### 3 **Availability of data and materials**

4 The data that support the findings of this study are available are available from the corresponding author  
5 upon request.

### 6 **Authors' contributions**

7 Wiklund P, Karhunen V and Järvelin M-R had full access to all of the data in the study and take responsibility  
8 for the integrity of the data and the accuracy of the data analysis. Concept and design: Wiklund P,  
9 Karhunen V, Rodriguez A, Relton C, and Järvelin M-R. Drafting of the work: Wiklund P, Karhunen V, Järvelin  
10 M-R. Critical revision of the manuscript for important intellectual content: All authors. Statistical analysis:  
11 Karhunen V., Richmond RC., Rezwan FI.

### 12 **Ethics approval and consent to participate**

13 All studies were approved by their local ethics committees and participants or their guardians provided  
14 written consent prior to the study.

### 15 **Consent for publication**

16 Not applicable.

### 17 **Competing interests**

18 The authors declare no competing interests.

19

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21

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27

## 1 **Figure legends**

2 Figure 1. Study design and analytical flow of the study. NFBC = Northern Finland Birth Cohort, ALSPAC =  
3 Avon Longitudinal Study of Parents and Children (m=mothers) and (c=children), IWBC = Isle of Wight Birth  
4 Cohort, EWAS = epigenome-wide association study. †CpG sites identified previously in cord blood of  
5 newborns exposed to maternal smoking in utero (20). \*Methylation data for persistence analysis.

6 Figure 2. Comparison of meta-analysis effect size estimates and their 95 % confidence intervals in all  
7 participants (x-axis) and never-smokers (y-axis) for the 36 top CpG sites. All effect size estimates are  
8 adjusted for study-specific covariates as necessary and meta-analyzed using inverse-variance weighted  
9 fixed-effects model.

10 Figure 3. Visualization of the dose-response effect of the intensity of maternal smoking in pregnancy (x-  
11 axis) on offspring blood DNA methylation (y-axis) for top four CpG sites in four gene regions (*AHRR*,  
12 *CYP1A1*, *MYO1G*, *GFI1*). Prediction estimates and their 95 % confidence intervals plotted based on  
13 generalized additive mixed models, with other covariates (offspring sex, body mass index, smoking status,  
14 population stratification and technical covariates) set at their mean (continuous variables) or mode  
15 (categorical variables). The density plots represent the distribution of the cigarettes smoked per day in  
16 pregnancy. The plots are truncated at five cigarettes per day in pregnancy (containing 94 % of full data).

17 Figure 4. Longitudinal analysis of association between exposure to maternal smoking and offspring blood  
18 DNA methylation. Effect size estimates (adjusted for study-specific covariates and meta-analyzed using  
19 inverse-variance weighted fixed-effects model) and their 95 % confidence intervals at age 30-31 years (red)  
20 and age 46-48 years (blue) for top CpG sites and *P* values for the test of equality of the effect size  
21 estimates.

22 Figure 5. Mediation analysis examining the indirect effect of maternal smoking during pregnancy on Bipolar  
23 II Scale (A) and Hypomanic personality scale (B) through differential methylation of cg25189904 in *GNG12*.  
24 Data are shown as beta estimate for effect size and 95 % confidence intervals.

1 Figure 6. A mediation model for the association between maternal smoking and offspring disease  
2 outcomes.  $\beta_a$  represent the effect estimate for smoking on DNA methylation (CpG = maternal smoking +  
3 covariates);  $\beta_b$  represents the effect estimate for CpG on disease (disease = CpG + covariates);  $\beta_c$  represent  
4 the direct effect (no mediation) estimate for maternal smoking on disease (disease = maternal smoking +  
5 covariates;  $\beta_c$  represents the total effect estimate on disease (disease = maternal smoking + covariates +  
6 CpG).

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1 **Table 1.** Association of exposure to maternal smoking during pregnancy and offspring peripheral blood DNA Methylation

CpG	Chr	Position	Gene	$\beta$ (SE)	P
cg04180046	7	45002736	<i>MYO1G</i>	0.045 (0.003)	2.60E-54
cg12803068	7	45002919	<i>MYO1G</i>	0.077 (0.005)	5.70E-45
cg22132788	7	45002486	<i>MYO1G</i>	0.045 (0.003)	6.80E-38
cg19089201	7	45002287	<i>MYO1G</i>	0.035 (0.003)	1.20E-31
cg05549655	15	75019143	<i>CYP1A1</i>	0.010 (0.001)	1.00E-28
cg25949550	7	145814306	<i>CNTNAP2</i>	-0.007 (0.001)	5.40E-27
cg18493761	11	125386885		0.037 (0.004)	7.30E-21
cg15507334	10	14372913	<i>FRMD4A</i>	0.024 (0.003)	1.80E-20
cg14179389	1	92947961	<i>GFI1</i>	-0.028 (0.003)	5.00E-20
cg00253658	16	54210496		0.037 (0.004)	3.40E-19
cg17924476	5	323794	<i>AHRR</i>	0.024 (0.003)	4.40E-19
cg13570656	15	75019196	<i>CYP1A1</i>	0.036 (0.004)	7.40E-19
cg11813497	10	14372879	<i>FRMD4A</i>	0.026 (0.003)	9.30E-19
cg22549041	15	75019251	<i>CYP1A1</i>	0.041 (0.005)	2.60E-18
cg05575921	5	373378	<i>AHRR</i>	-0.019 (0.002)	3.50E-18
cg12101586	15	75019203	<i>CYP1A1</i>	0.032 (0.004)	1.20E-17
cg18092474	15	75019302	<i>CYP1A1</i>	0.036 (0.004)	1.50E-17
cg11924019	15	75019283	<i>CYP1A1</i>	0.013 (0.002)	2.90E-16
cg25464840	10	14372910	<i>FRMD4A</i>	0.019 (0.002)	9.30E-16
cg00213123	15	75019070	<i>CYP1A1</i>	0.012 (0.002)	7.80E-15
cg11207515	7	146904205	<i>CNTNAP2</i>	-0.023 (0.003)	1.00E-13
cg14157435	2	206628692	<i>NRP2</i>	-0.046 (0.006)	1.10E-13
cg05348875	2	206628625	<i>NRP2</i>	-0.040 (0.006)	7.60E-13
cg01952185	5	134813213		0.019 (0.003)	3.30E-12
cg22308949	2	206628553	<i>NRP2</i>	-0.022 (0.003)	4.40E-12
cg05204104	2	235403141	<i>ARL4C</i>	0.017 (0.003)	4.80E-12
cg11641006	2	235213874		0.017 (0.003)	7.90E-12
cg09935388	1	92947588	<i>GFI1</i>	-0.029 (0.004)	1.10E-11
cg05697249	11	111789693	<i>C11orf52</i>	0.015 (0.002)	1.30E-11
cg21161138	5	399360	<i>AHRR</i>	-0.013 (0.002)	1.50E-11
cg26681628	16	54210550		0.018 (0.003)	1.80E-11
cg11025974	2	152830521	<i>CACNB4</i>	0.016 (0.002)	9.10E-11
cg15016771	2	235403218	<i>ARL4C</i>	0.008 (0.001)	1.00E-10
cg11429111	5	134813329		0.014 (0.002)	1.30E-10
cg25189904	1	68299493	<i>GNG12</i>	-0.020 (0.003)	1.30E-10
cg06758350	21	36259460	<i>RUNX1</i>	0.030 (0.005)	1.70E-10
cg17852385	15	75019188	<i>CYP1A1</i>	0.010 (0.002)	2.60E-10
cg01664727	21	36258423	<i>RUNX1</i>	0.027 (0.004)	2.60E-10
cg20344448	10	14372431	<i>FRMD4A</i>	0.014 (0.002)	2.80E-10
cg00794911	6	166260532		-0.012 (0.002)	5.20E-10
cg22698744	21	36263808	<i>RUNX1</i>	0.023 (0.004)	7.30E-10
cg03142697	21	36258497	<i>RUNX1</i>	0.016 (0.003)	8.10E-10

cg14563637	9	98931801		0.016 (0.003)	1.20E-09
cg15091747	21	36262896	<i>RUNX1</i>	0.014 (0.002)	1.20E-09
cg12984635	19	44032076	<i>ETHE1</i>	0.015 (0.002)	2.20E-09
cg01825213	9	98979965		0.019 (0.003)	2.50E-09
cg12477880	21	36259241	<i>RUNX1</i>	0.038 (0.006)	2.80E-09
cg17199018	8	28206278	<i>ZNF395</i>	-0.017 (0.003)	3.90E-09
cg00174179	3	49450293	<i>RHOA;TCTA</i>	-0.006 (0.001)	7.00E-09
cg15578140	7	147718109	<i>MIR548F3;CNTNAP2</i>	0.011 (0.002)	7.50E-09
cg14540913	9	132458514	<i>PRRX2</i>	0.013 (0.002)	7.60E-09
cg18132363	6	166260572		-0.019 (0.003)	7.60E-09
cg21253335	5	87835928		0.017 (0.003)	1.30E-08
cg25879142	7	4671391		0.018 (0.003)	1.80E-08
cg11845417	11	111789613	<i>C11orf52</i>	0.011 (0.002)	2.40E-08
cg20117519	7	8429907		0.022 (0.004)	2.40E-08
cg05783384	2	218843735		0.021 (0.004)	2.60E-08
cg13822849	9	137999757	<i>OLFM1</i>	0.007 (0.001)	2.90E-08
cg16449012	4	17781880	<i>FAM184B</i>	0.014 (0.002)	3.10E-08
cg05634495	6	122364658		0.016 (0.003)	3.10E-08
cg08644678	4	17711202	<i>FAM184B</i>	0.009 (0.002)	3.10E-08
cg13834112	15	90361639		0.013 (0.002)	3.60E-08
cg06635952	2	70025869	<i>ANXA4</i>	0.011 (0.002)	5.50E-08
cg14485097	7	4671479		0.016 (0.003)	7.40E-08
cg04598670	7	68697651		-0.019 (0.003)	8.30E-08
cg03252786	11	125106056	<i>PKNOX2</i>	0.006 (0.001)	8.70E-08
cg04749740	2	65935124		0.015 (0.003)	9.20E-08
cg15325070	1	2792704		0.014 (0.003)	9.20E-08
cg04358214	16	67143304	<i>C16orf70</i>	0.022 (0.004)	9.60E-08

1 The analyses were conducted separately in each participating cohort adjusted for study-specific covariates  
2 as necessary, and combined using inverse-variance weighted fixed-effects meta-analysis. Chr =  
3 chromosome;  $\beta$  = effect size estimate, SE = standard error.

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1 **Table 2.** Association results for the leading CpG sites from each locus selected for the sensitivity and downstream analyses.

CpG	Chr	Gene	Meta-analysis			Sensitivity and downstream analyses								
			(5 studies) N = 2821			Never-smokers N = 1298			Paternal smoking N = 1774			Dose-response analysis N = 1134		
			$\beta$	SE	P-value	$\beta$	SE	P-value	$\beta$	SE	P-value	$\beta$	SE	P-value
cg15325070	1		0.014	0.003	9.23E-08	0.009	0.004	1.92E-02	0.000	0.003	9.16E-01	0.168	0.034	6.81E-07
cg25189904	1	<i>GNG12</i>	-0.020	0.003	1.28E-10	-0.018	0.004	1.87E-05	-0.004	0.003	1.52E-01	-0.146	0.032	5.85E-06
cg14179389	1	<i>GFI1</i>	-0.028	0.003	5.00E-20	-0.027	0.004	1.07E-09	-0.004	0.003	1.77E-01	-0.214	0.031	4.92E-12
cg04749740	2		0.015	0.003	9.21E-08	0.015	0.004	9.64E-04	0.004	0.003	1.03E-01	0.061	0.034	6.98E-02
cg06635952	2	<i>ANXA4</i>	0.011	0.002	5.54E-08	0.011	0.003	1.88E-04	0.001	0.002	9.53E-01	0.073	0.029	1.12E-02
cg11025974	2	<i>CACNB4</i>	0.016	0.002	9.06E-11	0.018	0.004	1.57E-07	-0.001	0.002	6.41E-01	0.169	0.033	2.19E-07
cg14157435	2	<i>NRP2</i>	-0.046	0.006	1.15E-13	-0.042	0.008	6.01E-07	-0.014	0.006	2.80E-02	-0.201	0.033	9.33E-10
cg05783384	2		0.021	0.004	2.59E-08	0.014	0.005	1.15E-02	0.003	0.004	3.92E-01	0.097	0.033	3.26E-03
cg05204104	2	<i>ARL4C</i>	0.017	0.003	4.81E-12	0.016	0.004	6.02E-05	0.006	0.003	7.96E-02	0.036	0.033	2.73E-01
cg00174179	3	<i>RHOA</i>	-0.006	0.001	7.01E-09	-0.006	0.002	3.22E-04	-0.002	0.001	5.21E-02	-0.104	0.029	3.89E-04
cg16449012	4	<i>FAM184B</i>	0.014	0.002	3.10E-08	0.021	0.004	4.42E-08	0.003	0.002	1.62E-01	0.119	0.03	7.81E-05
cg05575921	5	<i>AHRR</i>	-0.019	0.002	3.53E-18	-0.011	0.003	1.19E-04	-0.008	0.003	2.75E-03	-0.134	0.027	5.91E-07
cg21253335	5		0.017	0.003	1.25E-08	0.014	0.004	1.12E-03	0.004	0.003	1.17E-01	0.108	0.033	1.04E-03
cg01952185	5		0.019	0.003	3.31E-12	0.017	0.004	5.81E-05	0.005	0.003	4.49E-02	0.128	0.03	1.67E-05
cg05634495	6		0.016	0.003	3.14E-08	0.014	0.005	2.54E-03	0.002	0.003	5.00E-01	0.088	0.033	6.86E-03

cg00794911	6		-0.012	0.002	5.20E-10	-0.007	0.003	1.25E-02	-0.005	0.002	1.15E-02	-0.115	0.034	6.42E-04
cg25879142	7		0.018	0.003	1.79E-08	0.013	0.004	3.05E-03	0.003	0.003	3.73E-01	0.121	0.031	1.23E-04
cg20117519	7		0.022	0.004	2.39E-08	0.022	0.006	1.70E-04	0.007	0.004	4.08E-02	0.112	0.033	6.72E-04
cg19089201	7	<i>MYO1G</i>	0.035	0.003	1.18E-31	0.03	0.004	5.75E-12	0.007	0.003	1.51E-02	0.225	0.032	2.24E-12
cg04598670	7		-0.019	0.003	8.33E-08	-0.016	0.005	4.54E-03	-0.007	0.003	3.95E-02	-0.178	0.034	1.54E-07
cg25949550	7	<i>CNTNAP2</i>	-0.007	0.001	5.41E-27	-0.007	0.001	1.45E-14	-0.002	0.001	6.57E-03	-0.157	0.022	1.65E-12
cg11207515	7	<i>CNTNAP2</i>	-0.023	0.003	1.03E-13	-0.02	0.004	2.75E-06	-0.011	0.003	2.46E-04	-0.145	0.032	7.02E-06
cg15578140	7	<i>MIR548F3</i>	0.011	0.002	7.54E-09	0.014	0.003	7.89E-06	-0.001	0.002	5.70E-01	0.152	0.031	1.12E-06
cg17199018	8	<i>ZNF395</i>	-0.017	0.003	3.85E-09	-0.016	0.004	7.10E-05	-0.003	0.003	3.10E-01	-0.121	0.033	2.63E-04
cg14563637	9		0.016	0.003	1.19E-09	0.016	0.004	9.26E-05	0.003	0.002	1.69E-01	0.092	0.031	3.26E-03
cg14540913	9	<i>PRRX2</i>	0.013	0.002	7.57E-09	0.015	0.003	1.81E-05	0.001	0.002	5.79E-01	0.148	0.032	2.62E-06
cg13822849	9	<i>OLFM1</i>	0.007	0.001	2.87E-08	0.007	0.002	5.84E-05	0.003	0.002	2.90E-02	0.126	0.03	3.24E-05
cg11813497	10	<i>FRMD4A</i>	0.026	0.003	9.27E-19	0.022	0.004	1.46E-07	-0.001	0.003	6.26E-01	0.117	0.032	2.81E-04
cg05697249	11	<i>C11orf52</i>	0.015	0.002	1.30E-11	0.015	0.003	8.68E-06	0.003	0.002	1.54E-01	0.136	0.035	8.25E-05
cg18493761	11		0.037	0.004	7.30E-21	0.032	0.006	4.99E-08	0.008	0.004	2.60E-02	0.154	0.034	4.82E-06
cg05549655	15	<i>CYP1A1</i>	0.010	0.001	1.04E-28	0.009	0.001	6.03E-13	0.002	0.001	4.28E-02	0.23	0.028	1.92E-16
cg13834112	15		0.013	0.002	3.55E-08	0.018	0.004	6.39E-07	0.001	0.002	8.10E-01	0.12	0.031	1.24E-04
cg00253658	16		0.037	0.004	3.44E-19	0.032	0.006	6.54E-08	0.005	0.005	2.81E-01	0.166	0.033	4.83E-07
cg04358214	16	<i>C16orf70</i>	0.022	0.004	9.56E-08	0.021	0.006	5.93E-04	0.005	0.004	2.42E-01	0.09	0.031	3.94E-03

cg12984635	19	<i>ETHE1</i>	0.015	0.002	2.17E-09	0.014	0.003	5.87E-05	0.001	0.002	7.33E-01	0.116	0.031	1.49E-04
cg06758350	21	<i>RUNX1</i>	0.030	0.005	1.68E-10	0.016	0.007	2.03E-02	0.002	0.005	7.09E-01	0.14	0.034	3.92E-05

1 The analyses were conducted separately in each participating cohort, adjusted for study-specific covariates as necessary, and combined using inverse-  
2 variance weighted fixed-effects meta-analysis. The effect size estimates for the dose-response analysis represent the difference in blood DNA methylation  
3 (in standard deviation units) per three additional cigarettes smoked per day in pregnancy. Chr = chromosome;  $\beta$  = effect size estimate, SE = standard error.

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1 **Table 3.** Mendelian randomization analysis of top differentially methylated CpGs tested against 106  
 2 diseases.

exposure	gene	disease	$\beta$	se	pval	pval FDR	unit
cg15578140	<i>MIR548F3</i>	Inflammatory bowel disease	-0.104	0.018	3.73E-09	2.54E-07	log odds
cg09935388	<i>GFI1</i>	Inflammatory bowel disease	-0.152	0.034	7.27E-06	0.000218	log odds
cg04598670	unknown	Inflammatory bowel disease	-0.410	0.091	7.27E-06	0.000524	log odds
cg09935388	<i>GFI1</i>	Crohn's disease	-0.162	0.040	4.74E-05	0.000712	log odds
cg04598670	unknown	Crohn's disease	-0.439	0.108	4.74E-05	0.001708	log odds
cg09935388	<i>GFI1</i>	Ulcerative colitis	-0.160	0.042	0.000147	0.001467	log odds
cg04598670	unknown	Ulcerative colitis	-0.433	0.114	0.000147	0.003521	log odds
cg25189904	<i>GNG12</i>	Schizophrenia	-0.222	0.053	3.37E-05	0.001819	log odds

3 Data are given as beta coefficients and their standard errors. Only significant associations ( $P_{FDR} <$   
 4 0.05) are shown. *GFI1* = Growth Factor Independent Protein 1; *MIR548F3* = microRNA 548f-3;  
 5 *GNG12* = Guanine Nucleotide Binding Protein Gamma 12. FDR = false discovery rate.

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1 **Table 4.** Data availability in each study for different analyses.

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Study	Mean age at methylation data collection (years)	Association results for cord-blood-methylation associated CpG sites	Sensitivity and downstream analyses for top CpG sites			
			Never-smokers	Paternal smoking	Amount of cigarettes smoked in pregnancy	Longitudinal methylation data (mean age in years at 2 <sup>nd</sup> time point)
NFBC1986	16	Yes	Yes	Yes	Yes	No
NFBC1966	31	Yes	Yes	Yes	Yes	Yes (46)
ALSPAC mothers	30	Yes	Yes	No	No	Yes (48)
ALSPAC children	17	Yes	Yes	Yes	No	No
IOWBC	18	Yes	Yes	No	No	No

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